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Corticosterone, prolactin and egg neglect behavior in relation to mercury and legacy POPs in a long-lived Antarctic bird



S. Tartu ^{a,*}, F. Angelier ^a, J.C. Wingfield ^b, P. Bustamante ^c, P. Labadie ^{d,e}, H. Budzinski ^{d,e}, H. Weimerskirch ^a, J.O. Bustnes ^f, O. Chastel ^a

- ^a UMR 7372, CNRS-Université de La Rochelle, Villiers-en-bois, France
- ^b Department of Neurobiology, Physiology and Behavior, University of CA, Davis, USA
- ^c Littoral Environnement et Sociétés (LIENSs), UMR 7266 CNRS-Université de la Rochelle, La Rochelle, France
- ^d Université de Bordeaux, EPOC/LPTC, UMR 5805, F-33400 Talence, France
- e CNRS, EPOC/LPTC, UMR 5805, F-33400 Talence, France
- f Norwegian Institute for Nature Research, FRAM High North Research Centre on Climate and the Environment, NO-9296 Tromsø, Norway

HIGHLIGHTS

- We measured legacy POPs and Hg in the blood of adult snow petrels from Antarctica.
- · POPs or Hg concentrations were not related to age.
- Stress-induced CORT concentrations were positively related to POP concentrations.
- · Stress-induced PRL concentrations were negatively related to Hg concentrations.
- Hg concentrations were higher in males that neglected their egg.

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ABSTRACT

Seabirds often have high loads of contaminants. These contaminants have endocrine disrupting properties but their relationships with some endocrine mechanisms are still poorly investigated in free-living organisms. This is the case for the stress response which shifts energy investment away from reproduction and redirects it towards survival. In birds, this stress response is achieved through a release of corticosterone and is also accompanied by a decrease in circulating prolactin, an anterior pituitary hormone widely involved in regulating parental cares. We measured blood concentrations of some legacy persistent organic pollutants (POPs) and mercury (Hg) and examined their relationships with the corticosterone and prolactin responses of known-age (9–46 years old) incubating snow petrels (Pagodroma nivea) to a standardized capture/handling stress protocol. In this Antarctic seabird, we also investigated whether high contaminant burden correlates with a higher occurrence of egg neglect, a frequently observed behavior in snow petrels. POPs and Hg were unrelated to age. Stress-induced corticosterone concentrations were positively related to POPs in both sexes, and stress-induced prolactin concentrations were negatively related to Hg in males. Egg-neglect behavior was not related to POPs burden, but males with higher Hg concentrations were more likely to neglect their egg. This suggests that in birds, relationships between age and contaminants are complex and that even low to moderate concentrations of POPs and Hg are significantly related to hormonal secretion. In this Antarctic species, exposure to legacy POPs and Hg could make individuals more susceptible to environmental stressors such as ongoing disturbances in Polar Regions. © 2014 Elsevier B.V. All rights reserved.

1. Introduction

The parental phase is energy-demanding (Drent and Daan 1980) and individuals adopt different life-history strategies in order to cope with environmental stressors such as food shortage, predation or poor

* Corresponding author.

E-mail address: tartu.sabrina@gmail.com (S. Tartu).

weather. In extreme environments, such as Polar Regions, animals often experience harsh and unpredictable environmental conditions, and as a result long-lived organisms such as seabirds may refrain from breeding or desert their brood (e.g. Angelier et al., 2007; Goutte et al., 2011a). At the physiological level, the release of glucocorticoid hormones (cortisol, corticosterone: CORT) during stressful events triggers physiological and behavioral adjustments that shift energy investment away from reproduction and redirects it towards self-preservation and

hence survival (Angelier and Wingfield, 2013; Ricklefs and Wikelski, 2002; Wingfield and Sapolsky, 2003). Stress hormones have therefore a strong connection to fitness traits such as breeding success, individual quality and survival (Angelier et al., 2009a, 2010; Bonier et al., 2009; Bókony et al., 2009; Breuner et al., 2008; Goutte et al., 2010, 2011b; Kitaysky et al., 1999). Additionally, the hormone prolactin (PRL) can also mediate this life-history trade-off between reproduction and survival in free-living birds (reviewed in Angelier and Chastel, 2009). The release of this anterior pituitary hormone stimulates and facilitates parental behavior such as egg incubation and brood provisioning (Buntin, 1996). In response to acute stress, circulating PRL levels has been shown to decrease in several bird species (Angelier et al., 2013; Chastel et al., 2005), and this could ultimately trigger nest desertion if PRL levels remain low during a prolonged period (Angelier et al., 2007, 2009b; Angelier and Chastel, 2009; Heidinger et al., 2010). Importantly, this decrease in PRL levels varies between individuals and lifehistory stages, suggesting that birds can attenuate their PRL response to acute stress to ensure that reproduction is not inhibited when the fitness value of the current reproductive event is high (the 'brood value hypothesis'; Bókony et al., 2009; Lendvai et al., 2007). Thus, both CORT and PRL are very likely to mediate parental effort and parental investment in birds (Angelier et al., 2007, 2009b, 2013; Chastel et al., 2005; Criscuolo et al., 2005; Groscolas et al., 2008; Koch et al., 2004) and any disruption of these major endocrine cascades may alter the ability of an individual to adjust reproductive decisions to environmental conditions (Jenssen, 2005; Tartu et al., 2013).

In addition to extreme environmental conditions, climate change and anthropogenic disturbances (Clarke and Harris, 2003; Moline et al., 2008; Smetacek and Nicol, 2005), polar species are subjected to environmental pollution. Indeed, despite their remote location, polar areas are the fall-out region of contaminants which undergo long range transport such as persistent organic pollutants (POPs) and heavy metals (e.g. mercury: Hg). Indeed, because of climate characteristics, contaminants accumulate in the polar environment, where they may be bio-accumulated and for some compounds bio-magnified (Bargagli, 2008; Gordeev, 2002; Risebrough et al., 1976; Wania and Mackay, 1996; Wania, 2003). Moreover, long-lived organisms are thought to be highly sensitive to contaminants (Rowe, 2008), but there are surprisingly few data on the effect of age on contaminant levels, and it is not clear if seabirds accumulate POPs with increasing age (Bustnes et al., 2003).

Marine apex predators, such as seabirds, are particularly exposed (Gabrielsen, 2007; Rowe, 2008; van den Brink, 1997) and several studies have reported breeding impairments in highly polluted seabirds (Bustnes et al., 2001, 2003, 2007; Tartu et al., 2013; Verboven et al., 2009). Such breeding impairments could originate from the ability of contaminants to act as endocrine disruptors and thus, to alter the functioning of endocrine axes (Guillette and Gunderson, 2001; Ottinger et al., 2002, 2013; Tan et al., 2009; Tartu et al., 2013, 2014; Tyler et al., 1998). Experimental studies have documented some effects of chemicals on glucocorticoids (Love et al., 2003; Odermatt and Gumy, 2008), but the effects of contaminants on stress hormones in free-living organisms such as seabirds have rarely been studied (Bergman et al., 2013; Nordstad et al., 2012; Tartu et al., 2014; Verboven et al., 2010). It is therefore difficult to draw a general pattern of the relationships between contaminants and stress hormones.

Regarding the effects of contaminants on PRL, the knowledge is even poorer and only one study (Verreault et al., 2008) has addressed relationships between PRL secretion and POPs. In glaucous gulls (*Larus hyperboreus*) baseline PRL levels and the rate of decrease in PRL levels tended to vary negatively with organohalogen contaminants in males only (Verreault et al., 2008). Furthermore, numerous compounds are potential environmental contaminants (e.g. heavy metals, POPs), which may have different effects on hormones of the hypothalamopituitary–adrenal (HPA) axis, such as CORT but also PRL secretion. There is thus a need to determine whether different environmental

contaminants can disrupt these hormones in free-living organisms. The aim of this study was to investigate the potential roles of environmental contaminants such as Hg and some legacy POPs (i.e. polychlorinated biphenyls: PCBs; organochlorine pesticides: OCPs; and polybrominated diphenyl ethers: PBDEs) on two major endocrine mechanisms: stress hormones from the HPA axis: CORT, and a key pituitary 'parental hormone': PRL. The snow petrel (Pagodroma nivea) a contaminated Antarctic seabird (Xie et al., 2008; Corsolini et al., 2011; Goutte et al., 2013) provides an ideal species to address these questions. In this long-lived species (until ~50 years old, Chastel et al., 1993), CORT and PRL responses to acute stress are modulated in relation to parental investment and incubation commitment (Angelier et al., 2007; Goutte et al., 2011c). For example, low stress-induced PRL levels are associated with a high probability of egg neglect, a frequently observed behavior in snow petrels (Angelier et al., 2007). Further, thanks to an exceptional long-term banding survey (1964-present; Barbraud and Weimerskirch, 2001; Chastel et al., 1993), many snow petrels are of known age, making it possible to address the effect of age on contaminant burden.

In that context, we investigated if POPs and/or Hg concentrations were related to 1) age, 2) CORT and/or PRL secretion and 3) parenting through egg-neglect behavior. We predicted that POPs and/or Hg: 1) would increase with increasing age as a result of bio-accumulation; 2) would increase CORT and decrease PRL secretion; 3) would be higher in individuals that neglected their egg.

2. Materials and methods

2.1. Ethics statement

Animals were cared for in accordance with the guidelines of the ethics committee of the Institut Polaire Français Paul Emile Victor (IPEV) that specifically approved this study (Program no. 109, H. Weimerskirch).

2.2. Study site, blood sampling and body-condition

Snow petrels are Antarctic seabirds with a delayed sexual maturity (~10 years of age), a low fecundity (one egg per clutch and a maximum of one clutch per year) and a long lifespan (~50 years old) (Chastel et al., 1993). Adult males and females were handled during the 2010 late incubation period (8-21 January). A total of 49 birds (27 males and 22 females) were caught in 49 different nests and age was known for 47 of them (9–46 years old). Birds were captured by hand and were then bled according to the standardized capture/restraint stress protocol described by Wingfield (1994). Immediately after capture (i.e. within 3 min), an initial blood sample (300 µl) was collected from the alar vein with a 1-ml heparinized syringe and a 25-gauge needle. These initial blood samples were considered to reflect baseline levels of CORT and PRL (Chastel et al., 2005; Romero and Reed, 2005; thereafter called 'baseline' sample). After collection of the initial blood samples birds were placed into cloth bags, and a subsequent sample (300 µl) was collected 30 min after capture (thereafter called 'stress-induced' sample). During handling of the adult birds, their eggs were covered with cotton and kept warm. After these blood samples, each bird was put back in its nest. Snow petrels are tame and usually resume parental duties as soon as returned to their nest (e.g. Angelier et al., 2007). After this acute stress protocol, petrels were left undisturbed at their nest for 20 min and were then captured again and blood sampled within 3 min of recapture (thereafter called 'post-stress' sample) to monitor how quickly hormone levels may return to baseline after a stressor. This blood sample was taken before CORT and PRL concentrations returned to normal, allowing us to effectively monitor the stress recovery. All birds were weighed to the nearest 2 g using a spring balance and their skull length (head + bill) was measured to the nearest 0.5 mm. Body condition index (thereafter 'body condition') was calculated as the residuals between body mass and skull length (regression: $F_{1.47} = 20.28$,

p < 0.001, $R^2 = 0.35$). After capture and blood sample, each nest was monitored twice a day until the manipulated petrel was relieved by its mate. We were therefore able to know whether a bird neglected its egg during the incubation bout following capture/restraint stress protocol (thereafter called 'egg neglect behavior'). Leaving eggs unattended temporarily is common in Procellariiform birds (Boersma and Wheelwright, 1979; Chaurand and Weimerskirch, 1994). Distant foraging and unpredictable weather increase the probability to delay an individual's returning to relieve its incubating partner (Boersma and Wheelwright, 1979). Eggs left unattended for a long period are less likely to hatch successfully (Boersma and Wheelwright, 1979; Angelier et al., 2007). In snow petrels were egg-neglect is often observed (Angelier et al, 2007), both parents incubate the single egg four bouts lasting ca. 4 to 8 days while the partner is feeding at sea (Ryan and Watkins, 1989). In two sampled birds, the egg was predated during the incubation bout following the capture/restraint stress protocol. Egg-neglect data were available for 47 birds.

2.3. Molecular sexing and hormone assay

Blood samples were centrifuged, and plasma was decanted and stored at -20 °C until assayed. After centrifugation, red cells were kept frozen for molecular sexing as well as for Hg determination. The sex was determined by polymerase chain reaction amplification of part of two highly conserved genes (CHD) present on the sex chromosomes at UMR 7372, CNRS-Université de La Rochelle, as detailed in Weimerskirch et al. (2005). Plasma concentrations of CORT were determined first by radioimmunoassay at UMR 7372, CNRS-Université de La Rochelle, as previously described (Lormée et al., 2003). Plasma concentrations of PRL were determined with the remaining plasma by a heterologous radioimmunoassay at UMR 7372, CNRS-Université de La Rochelle, as detailed in Cherel et al. (1994). The PRL assay has previously been validated in snow petrels (Angelier et al., 2007). All samples were run in one assay for both hormones. To measure intra-assay variation, we included 4 different referents 10 times in the CORT and PRL assays. From this, the intra-assay variation was 6.7% for total CORT and 7.8% for PRL. CORT and PRL concentrations were measured in baseline, stress-induced and post-stress samples.

2.4. Organic pollutants determination in plasma

POPs were measured in plasma samples collected from 15 females and 21 males only, since in 13 birds the remaining plasma volumes were too low. The targeted compounds included 7 indicator PCBs (CB-28, -52, -101, -118, -138, -153 and -180), 11 OCPs (HCB, Gamma HCH, Heptachlore, cis-chlordane, trans-nonachlor, 2,4' DDE, 4,4' DDE, 4,4' DDD, 2,4' DDT, 4,4' DDT and mirex) and two PBDE (BDE-47 and BDE-99). Certified solutions containing all analytes in isooctane at 2 ng·µl⁻¹ each were obtained from LGC Standards (Molsheim, France). To a plasma sample of 100 µl, internal standards (1 ng each) were added gravimetrically: CB-30, -103, -155 and -198 were used to quantify PCBs, p,p'-DDT-d8 was used to quantify OCPs and F-BDE-47 was used to quantify BDE-47 and BDE-99; standards were provided by either Dr. Ehrenstorfer GmbH, Cambridge Isotope Laboratory (via Cluzeau Info Labo, Sainte-Foy-La-Grande, France) or Chiron (via BCP Instruments, Irigny, France). POPs were extracted with 1 ml of pentane:dichloromethane (90:10; v/v); after centrifugation (2000 rpm, 2 min at 4 °C), the organic layer was collected and the operation was repeated. Both extracts were combined and purified on an acid silica gel column (40% H₂SO₄). After extract loading, analytes were eluted with 3 \times 5 ml of pentane/dichloromethane (90/10; v/v). Extracts were then concentrated using a RapidVap vacuum evaporation system from Labconco (Kansas City, MO, USA) to a volume of 1 ml and further concentrated under a gentle stream of nitrogen (40 °C) after addition of 100 µl of isooctane as solvent keeper. Octachloronaphtalene (1 ng) was finally added to determine the recovery rate for each internal standard, for each sample (68–108%). Final extracts were analyzed by gas chromatography coupled with electron capture detection (GC-ECD) as described elsewhere (Tapie et al., 2011).

Quality control consisted in the analysis of standard solutions (NIST SRM 2261 and SRM 2262) and of procedural blanks (clean and empty glass tubes treated like a sample, one blank for 8 samples). Recoveries for standard solutions ranged from 89 to 104% with standard deviations lower than 13% (n = 4). Chicken plasma samples (Sigma-Aldrich, St. Quentin Fallavier, France) spiked with all analytes (3 $ng \cdot g^{-1}$ each) were analyzed; the recovery rates were in the range 77-103% with coefficients of variation lower than 17% (n = 5), except for CB-52 (22%) and mirex (29%). POP concentrations were blank corrected and the detection limit (LoD) was set at two times the mean blank value; for analytes that were not detected in blanks, LoD was determined as the concentration with a signal to noise ratio of 3 in spiked chicken plasma samples. Overall, LoDs ranged from 0.03 to 0.34 ng·g⁻¹ wet weight (ww). Additionally, plasma total lipids were measured on an aliquot of 10 µl by the sulfo-phospho-vanillin (SPV) method for colorimetric determination (Frings et al., 1972).

2.5. Hg determination in blood cells

Total Hg was measured as described in details in Bustamante et al. (2006). Briefly, from freeze-dried and powdered red blood cells (hereafter called 'blood') in an Advanced Hg Analyzer spectrophotometer (Altec AMA 254). At least two aliquots ranging from 5 to 10 mg were analyzed for each individual and quality assessment was measured by repeated analyses of certified reference material TORT-2 (lobster hepatopancreas, NRCC; certified value $0.27 \pm 0.06 \, \mu g \cdot g^{-1}$; with recoveries of 98 to 102%) and blanks, empty sample container, run every 20 samples. Hg concentrations are expressed in $\mu g \cdot g^{-1}$ dry weight (dw).

2.6. Statistical analyses

All analyses were performed using R 2.13.1 (http://r-project.org/). We first tested inter-correlations between the different families of POPs detected by using linear models (LM). Second, we used generalized linear model (GLM) with normal errors and an identity link function to test whether POPs or Hg were influenced by sex, body-condition and age (dependent variable: POPs and Hg, independent factor and variables: sex, body-condition and age). Third, we tested whether CORT and PRL kinetics differed between male and females by using repeated measures GLM with the time of sampling (baseline, stress-induced and post-stress levels) as the repeated measures; (dependent variable: CORT and PRL concentrations, independent factors: sex, time and their interaction). Fourth, we tested whether CORT concentration (baseline, stress-induced and post-stress) was related to POPs and Hg (dependent variable: CORT, independent factor and variables: POPs, sex, age, Hg, body-condition and their interaction with sex). For PRL we analyzed males and females separately (dependent variable: PRL, independent variables: POPs, age, Hg, body-condition) because in incubating snow petrels, females bear higher PRL concentrations than males (Angelier et al., 2007). Finally, we tested if the probability of neglecting the egg was related to POPs and Hg in males and females separately (dependent variable: egg neglect; independent variables: POPs, age, Hg, bodycondition). To test the relationships between contaminants and eggneglect behavior (yes or no) we used GLM with binomial error and logit link. Dependent continuous variables were previously tested for normality with a Shapiro-Wilk test and were log-transformed when necessary. Selected models were then checked for assumptions, that is, constancy of variance and residual normality. We performed all our model selection starting from the most general model that included all the variables/factors of interest and their interactions and we removed step by step the non-significant interactions, variables or factors. For POPs statistical analyses, concentrations below LoD were assigned LoD

value, and only compounds detected in at least 70% of the individuals were included into the sum of POPs (Noël et al., 2009).

3. Results

3.1. Contaminants: concentrations and compounds

Out of the 20 POP targeted compounds, 15 could be detected but the concentrations of four OCPs (cis-chlordane, trans-nonachlor, heptachlor and 2,4′-DDE) and one PBDE (BDE-99) were systematically below LoD. The most abundant compounds were the PCBs, with the PCBs 101 and 118 reaching the highest concentrations, closely followed by CB-138 and CB-153 (Table 1). Of the OCPs, HCB had the highest concentrations followed by 4,4′-DDE (Table 1). Only four PCBs (-CB101, -118, -138 and -153) and four OCPs (HCB, gamma HCH, 4,4′ DDE and 2,4′ DDT) were detected in at least 70% of the individuals and were thus included into the analyses. PCBs and OCPs were positively correlated (PCBs vs. OCPs: LM, $F_{1,34}=18.1,\,p<0.001,\,R^2=0.35$). Thus the global pollutant burden was described as a sum of POPs (hereafter 'POPs'). In incubating snow petrels, blood Hg averaged $1.91\pm0.75\,\mu g\cdot g^{-1}$ dw, specifically in males $1.94\pm0.77\,\mu g\cdot g^{-1}$ dw (range: 0.89–4.01) and in females $1.87\pm0.73\,\mu g\cdot g^{-1}$ dw (range: 0.74–3.70).

3.2. Relationship between contaminants, sex, body-condition and age

During the incubation period, POPs was not statistically different between male and female snow petrels (GLM, $F_{1,34}=3.21,\,p=0.082).$ Hg concentrations were not related to sex neither (GLM, $F_{1,47}=0.12,\,p=0.734).$ POPs and Hg concentration, respectively, were unrelated to the body-condition index (POPs: $F_{1,34}=1.05,\,p=0.313,$ interaction with sex: $F_{1,33}=2.18,\,p=0.149;\,Hg:\,F_{1,46}=3.30,\,p=0.076,$ interaction with sex: $F_{1,45}=0.14,\,p=0.714).$ POPs were not related to age ($F_{1,34}=0.88,\,p=0.355,$ interaction with sex: $F_{1,33}=0.61,\,p=0.441),$ and neither were Hg concentrations ($F_{1,45}=2.05,\,p=0.159$ interaction with sex: $F_{1,44}=0.16,\,p=0.693).$ Finally, POPs and Hg were not related ($F_{1,34}=2.09,\,p=0.157).$

3.3. CORT and PRL kinetics: response and recovery to acute stress protocol

CORT concentrations significantly increased over 30 min from 4.7 \pm 3.4 to 37.8 \pm 8.6 ng·ml $^{-1}$ and then declined 20 min post-stress to 23.6 \pm 12.6 ng·ml $^{-1}$ (GLMM, time as factor, F_{2,94} = 269.98, p < 0.001), without sex difference (sex: F_{1,47} = 0.28, p = 0.600; time × sex interaction: F_{2,94} = 0.08, p = 0.926). PRL concentrations significantly decreased over time (GLMM, time as factor, F_{2,94} =

Table 1 Concentrations of plasma persistent organic pollutants and blood Hg $(\mu g \cdot g^{-1})$ of incubating female and male snow petrels. For POPs, concentrations are given in wet-weight $(ng \cdot g^{-1} ww)$ in the first rows and lipid-weight $(ng \cdot g^{-1} w)$ in the second rows.

| | Females | | | Males | | |
|------------|---------|--------------------|------------------|--------|--------------------|------------------|
| | Mean | Standard deviation | Range (min-max) | Mean | Standard deviation | Range (min-max) |
| Lipids (%) | 0.68 | 0.1 | (0.48-0.88) | 0.7 | 0.1 | (0.50-0.94) |
| CB-50/28 | 0.4 | 0.6 | (<0.10-2.1) | 1.2 | 2.5 | (<0.10-10.9) |
| | 65.2 | 92.1 | (<14.4-322.7) | 152.2 | 286.7 | (<14.4-1160.8) |
| CB-52 | 3.8 | 9.5 | (<0.04-36.6) | 10.0 | 16.3 | (<0.04-63.1) |
| | 617.1 | 1494.2 | (<5.2-5719.4) | 1300.2 | 1953.0 | (<5.2-6709.3) |
| CB-101 | 6.9 | 14.4 | (<0.02-57.4) | 24.0 | 40.5 | (<0.02-167.9) |
| | 1082.1 | 2251.5 | (<3.1-8976.5) | 3151.6 | 5019 | (<3.1-20,730.2) |
| CB-118 | 6.6 | 19.0 | (<0.04-73.9) | 22.7 | 38.4 | (<0.04-150.6) |
| | 1045 | 2960.4 | (<5.4–11,543.5) | 2963.4 | 4802.2 | (<5.4–18,587.5) |
| CB-138 | 2.8 | 6.8 | (<0.02-26.5) | 12.1 | 17.5 | (<0.02-52.4) |
| | 441.5 | 1067 | (<2.5-4136.9) | 1601.4 | 2261.5 | (<2.5-6469.2) |
| CB-153 | 2.6 | 6.3 | (<0.04-24.6) | 9.8 | 13.9 | (<0.04-44.5) |
| | 414.9 | 988.0 | (<6.0-3845.6) | 1297.4 | 1780.2 | (<6.0-5488.8) |
| CB-180 | 0.2 | 0.6 | (<0.02-2.2) | 1.1 | 1.6 | (<0.02-5.4) |
| | 37 | 88.4 | (<3.2-338.8) | 145.8 | 204.3 | (<3.2-666.0) |
| PCBs* | 12.6 | 28.0 | (1.6-110.7) | 47.0 | 72.0 | (1.8-270.2) |
| | 1975.5 | 4379.2 | (211.4–17,297.8) | 5394.6 | 8427 | (222.2-33,354.2) |
| НСВ | 1.3 | 1.0 | (<0.01-3.2) | 2.2 | 1.6 | (<0.01-4.8) |
| | 192.6 | 153.2 | (<1.2-395.0) | 296.6 | 223.9 | (<1.2-779.1) |
| Gamma HCH | 0.1 | 0.1 | (<0.02-0.3) | 0.2 | 0.1 | (<0.02-0.5) |
| | 16.9 | 19.8 | (<2.1-47.8) | 21.7 | 16.8 | (<2.1-56.5) |
| 4,4'-DDE | 0.6 | 1.0 | (<0.01-4.0) | 0.4 | 0.4 | (<0.01-1.2) |
| | 92.7 | 168 | (<1.7-669.3) | 53.9 | 51.9 | (<1.7-192.3) |
| 4,4'-DDD | 0.1 | 0.1 | (<0.05-0.4) | 0.1 | 0.2 | (<0.05-0.9) |
| | 8.9 | 17.5 | (<7.3-65.7) | 18.2 | 27.9 | (<7.3-114.7) |
| 2,4'-DDT | 0.3 | 0.7 | (<0.03-2.8) | 1.1 | 1.9 | (<0.03-6.6) |
| | 47.4 | 114.9 | (<4.6-433.5) | 155.9 | 266.8 | (<4.6-1018.7) |
| 4,4'-DDT | 0.3 | 0.6 | (<0.02-2.2) | 1.3 | 2.5 | (<0.02-9.0) |
| | 40.2 | 92.0 | (<2.7-337.2) | 176.3 | 350.8 | (<2.7-1388.9) |
| Mirex | 0.2 | 0.3 | (<0.02-1.2) | 0.6 | 1.0 | (<0.02-3.4) |
| | 26.8 | 49.3 | (<2.6-190.4) | 78.1 | 124.2 | (<2.6-420.5) |
| OCPs** | 2.3 | 1.8 | (0.3-6.2) | 3.9 | 2.6 | (0.3-10.4) |
| | 349.5 | 286.7 | (33.1-1027.7) | 531.9 | 375.3 | (43.6-1593.1) |
| BDE-47 | 0.1 | 0.2 | (<0.03-0.7) | 0.2 | 0.3 | (<0.03-0.9) |
| | 10.2 | 29.8 | (<4.3–115.4) | 27.4 | 35.5 | (<4.3-95.7) |
| POPs*** | 14.9 | 29.0 | (2.3–116.1) | 50.8 | 73.5 | (3.9–275.3) |
| | 2325.1 | 4533.2 | (244.5–18,134.5) | 5926.5 | 8645.2 | (456.1-33,985.0) |
| Hg | 1.8 | 0.7 | (0.7–3.7) | 1.9 | 0.8 | (0.9-4.0) |

^{*} PCBs = CB101 + CB138 + CB153 + CB180.

^{**} OCPs = HCB + -HCH + 4,4'-DDE + 2,4'-DDT.

^{***} POPs = PCBs + OCPs.

144.26, p < 0.001) and females had higher PRL concentrations than males (sex: $F_{1,47}=152.72$, p < 0.001; time × sex interaction: $F_{2,94}=3.79$, p = 0.026). In females, PRL decreased from 239.5 \pm 36.3 to 169.4 \pm 32.1 ng·ml⁻¹ after 30 min and until 165.4 \pm 30.8 ng·ml⁻¹ 20 min post-stress. In males PRL concentrations decreased from 139.5 \pm 30.7 to 90.9 \pm 24.7 over 30 min and they reached 83.4 \pm 24.0 ng·ml⁻¹ 20 min post-stress.

3.4. Relationships between contaminants and CORT concentrations

CORT absolute concentrations (baseline, stress-induced and poststress) were not related to sex, neither to age, Hg, body-condition and their interaction with sex (Table 2). POPs were not related to baseline CORT (Fig. 1A, Table 2), but increasing concentration of POPs was positively related to increasing concentration of stress-induced and post-stress CORT (Fig. 1B–C, Table 2).

3.5. Relationships between contaminants, PRL concentrations and egg-neglect

In females we did not find any relationship between PRL (baseline, stress-induced and post-stress) and POPs, age, Hg, or body-condition (p > 0.08 for all tests). In males baseline and post-stress PRL concentrations were not related to POPs, age, Hg, or body-condition (p > 0.07 for all tests), but increasing blood Hg concentration was related to decreasing stress-induced PRL concentration: i.e. after 30 min restraint the most contaminated males were less likely to maintain high concentrations of PRL (GLM, $F_{1,25} = 5.6$, p = 0.0263; Fig. 2). Eleven females and eight males were observed neglecting their egg, out of 21 and 26, respectively. Blood Hg concentration was higher in males that were more likely to neglect their egg (GLM, $\chi^2 = 5.4$, p = 0.019, Fig. 3) a

Table 2 Relationships between contaminants (POPs and Hg), sex, age, body-condition and interaction with sex as a function of CORT concentrations $(ng \cdot ml^{-1})$ in incubating snow petrels: a) baseline, b) stress-induced and c) post-stress. N is the number of birds of each variable. Degrees of freedom vary between measures of CORT because model selection was performed by starting from the most general model that included all the variables/ factors of interest and their interactions and we removed step by step the non-significant interactions, variables or factors.

| Dependent variables | N | Independent variables | Df | F | p-Value |
|------------------------|----|-----------------------------|------|------|---------|
| a) Baseline CORT | 36 | Log (POPs) | 1,27 | 0.14 | 0.715 |
| | 49 | Sex | 1,47 | 1.2 | 0.279 |
| | 47 | Age | 1,44 | 0.2 | 0.66 |
| | 49 | Hg | 1,42 | 0.09 | 0.767 |
| | 48 | Body condition | 1,39 | 1.73 | 0.195 |
| | 36 | $Log (POPs) \times Sex$ | 1,26 | 0.46 | 0.505 |
| | 47 | $Age \times Sex$ | 1,43 | 3.49 | 0.068 |
| | 49 | $Hg \times Sex$ | 1,41 | 3.29 | 0.077 |
| | 49 | Body condition × Sex | 1,38 | 0.05 | 0.819 |
| b) Stress-induced CORT | 36 | Log (POPs) | 1,34 | 6.1 | 0.019 |
| | 49 | Sex | 1,33 | 0.48 | 0.495 |
| | 47 | Age | 1,30 | 0.02 | 0.9 |
| | 49 | Hg | 1,28 | 0.01 | 0.915 |
| | 48 | Body condition | 1,31 | 2.82 | 0.103 |
| | 36 | $Log (POPs) \times Sex$ | 1,26 | 0 | 0.989 |
| | 47 | $Age \times Sex$ | 1,29 | 0.64 | 0.429 |
| | 49 | $Hg \times Sex$ | 1,27 | 0.21 | 0.653 |
| | 49 | Body condition \times Sex | 1,32 | 3.97 | 0.055 |
| d) Post-stress CORT | 36 | Log (POPs) | 1,34 | 5.17 | 0.029 |
| | 49 | Sex | 1,33 | 1.2 | 0.282 |
| | 47 | Age | 1,30 | 0.33 | 0.57 |
| | 49 | Hg | 1,27 | 0.03 | 0.863 |
| | 48 | Body condition | 1,31 | 1.37 | 0.25 |
| | 36 | $log (POPs) \times Sex$ | 1,32 | 1.51 | 0.228 |
| | 47 | $Age \times Sex$ | 1,29 | 1.34 | 0.257 |
| | 49 | $Hg \times Sex$ | 1,26 | 0 | 0.955 |
| | 49 | Body condition \times Sex | 1,28 | 0.18 | 0.676 |

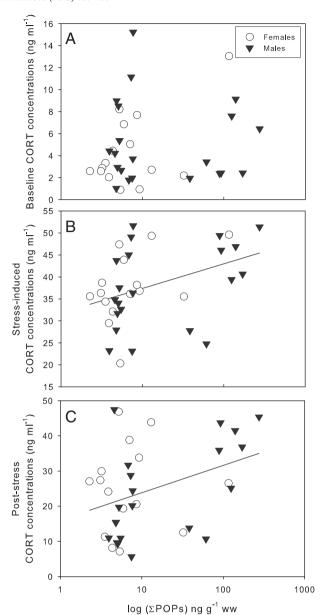


Fig. 1. Relationships between plasma POP concentrations ($\operatorname{ng} \cdot \operatorname{g}^{-1}$ wet weight) and plasma CORT concentrations ($\operatorname{ng} \cdot \operatorname{ml}^{-1}$) in incubating snow-petrels (A) baseline, (B) stress-induced and (C) post-stress. White circles denote females and black triangles denote males. The solid line refers to statistically significant linear regression.

relationship not found in females (GLM, $\chi^2 = 0.1$, p = 0.796, Fig. 3). Finally, egg neglect behavior was not related to POPs, age or bodycondition in any sex (p > 0.4 for all tests).

4. Discussion

The present study is the first to report plasma POP concentrations in the long-lived, Antarctic snow petrel. Firstly, there were no relationships between plasma POPs or blood Hg and age, suggesting that long-lived seabird are able to eliminate much of their contaminant burden. Secondly, POPs and Hg seem related to different hormonal pathways involved in reproductive decisions; POPs may disrupt the HPA axis whereas Hg was related to PRL secretion in males and consequently to egg-neglect behavior.

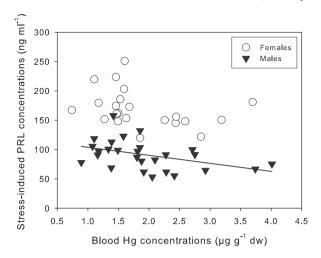


Fig. 2. Relationship between total blood Hg ($\mu g \cdot g^{-1}$ dry weight) and stress-induced plasma PRL concentrations ($ng \cdot ml^{-1}$) in incubating snow-petrels. White circles denote females and black triangles denote males. The solid line refers to statistically significant linear regression for males.

4.1. Contaminants and age

Although snow petrels are very long-lived and thus exposed to contaminants over many years, no evidence was found that contamination was age-related, neither for POPs nor Hg. POPs and Hg measured in blood (plasma and red blood cells, respectively) can be correlated to levels found in storage organs and also adipose tissues, in birds but also chelonians and humans (Henriksen et al., 1998; Henny et al., 2002; Pauwels et al., 2000; Wayland et al., 2001; Keller et al., 2004; van de Merwe et al., 2010; Szumiło et al., 2013). Thus, blood contaminant concentration may be a good proxy of contaminant burden in other organs.

The relationship between Hg and age in seabirds is often contradictory, for example liver Hg was found to decrease, increase or be unrelated to age (Furness and Hutton, 1979; Hutton, 1981; Thompson et al., 1991). For blood, the relationship between Hg contamination and age is also not clear: no relationship was found between age and Hg contamination (Gonzáles-Solís et al., 2002; Tavares et al., 2013) but in pre-breeding snow petrels and incubating cape petrels (*Daption capense*), a negative relationship was found between blood Hg and age. This relationship was, however more likely the result of an age-related change in feeding ecology. With regard to POPs, it seems that in seabirds, concentrations in different tissues and blood increase until a steady-state is reached, often before the age of breeding (Donaldson

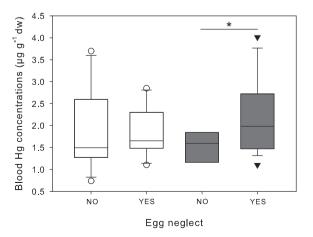


Fig. 3. Relationship between blood Hg ($\mu g \cdot g^{-1}$ dry weight) and egg-neglect behavior in incubating snow-petrels. The empty boxes denote females and the filled boxes denote males (*: p < 0.02).

et al., 1997; Newton et al., 1981; van den Brink et al., 1998), and for breeding birds, most studies have not observed any age-related POP accumulation (Bustnes et al., 2003; Newton et al., 1981). In this study, all snow petrels were breeders, and presumably they had already reached their steady-state levels.

Besides, seabirds can biotransform PCBs and eliminate POPs through their preen gland (Borgå et al., 2005; Henriksen et al., 1996; Solheim, 2010), in the same line Hg can be excreted through feather growth (Bearhop et al., 2000). These mechanisms could partially explain the lack of association between blood contaminants and increasing age. Also we have to remain cautious as we hypothesized that in snow petrels, as in other bird species, blood contaminants would represent levels in internal tissues (Henny et al., 2002; Szumiło et al., 2013; Wayland et al., 2001; Henriksen et al., 1998). However, we have no evidence for this relationship in snow petrels. Additionally, following food intake or lipid mobilization, contaminant levels in blood may fluctuate more than those in adipose tissues or liver and this could mask a hypothetical contaminant/age relationship.

4.2. POPs: concentrations and relationship with the CORT stress response

Very few studies have examined blood concentrations of POPs in adult Antarctic seabirds, and among the few studies, comparisons are made difficult due to different analytical methods. Indeed, POPs are often described as a "sum of compounds" and the compounds taken into the sum vary among studies. However, some OCPs are often reported individually since their detrimental effects have been well identified. This is the case of the HCB a relatively volatile compound (Calamari et al., 1991) principally used in fungicide formulations (Barber et al., 2005). Higher concentrations of this compound are commonly found in species restricted to the Antarctic region than those in temperate regions (e.g. van den Brink, 1997). HCB was the OCP with the highest concentrations in incubating snow petrels, but much lower than the concentrations found in other high-Antarctic species such as south polar skuas (Catharacta maccormicki) from Svarthamaren (71° 53′ S, 05° 10′ E) in Dronning Maud Land (Antarctica): i.e. HCB concentrations were 10-fold lower, and mirex concentrations 100-fold lower (Bustnes et al., 2006, 2007). These results could be the consequence of a different trophic level, toxicokinetic factors (e.g. metabolism, clearance rate), compound-specific physicochemical properties (Kow, half-life) or depend on the bio-availability of contaminants in the breeding area (Walker et al., 2012).

The Arctic is more contaminated by POPs than Antarctica (Bustnes et al., 2006; Choi et al., 2008), HCB concentrations in snow petrels were slightly lower than those measured in plasma of incubating black-legged kittiwakes (Rissa tridactyla) from Svalbard: 1.85 \pm 1.41 ng·g⁻¹ ww versus 2.5 \pm 0.44 ng·g⁻¹ ww respectively. In comparison, incubating glaucous gulls (Larus hyperboreus) had HCB concentrations in plasma much higher than snow petrels (Verreault et al., 2005): on average ~400 ng·g⁻¹ lipid weight whereas in snow petrels concentrations given in lipid weight average ~200 ng·g⁻¹. In both blacklegged kittiwakes and glaucous gulls there is evidence of CORT disruption by POPs (Nordstad et al., 2012; Verboven et al., 2010). Indeed, in both species increasing POP concentrations were related to higher baseline CORT concentrations and for male glaucous gulls, higher POP concentrations were related to decreasing stress-induced CORT concentrations. In the present study, increasing POPs were not related to baseline CORT, but to stress-induced and post-stress CORT concentrations. Hence, the most polluted birds released more CORT when subjected to a handling stress protocol, and those concentrations remained high 20 min post-stress. These results are in accordance with the recent finding that POPs, and especially PCBs are associated with a higher adrenocortical response to an acute stress in pre-laying female black-legged kittiwakes (Tartu et al., 2014). However, although post-stress CORT concentrations were admittedly higher in the most contaminated snow petrels they did not decrease more slowly than in less polluted birds,

indicating that negative feedback from CORT on the hypothalamus and the pituitary was functional.

One possible explanation for the over-release of CORT could be related to an increase of the number of adreno-corticotrophic-hormone (ACTH) receptors (ACTH-R) on the adrenals. ACTH is one of the few polypeptide hormones having a positive trophic effect on its own receptors (Beuschlein et al., 2001; Penhoat et al., 1989). Thus, an increase of ACTH-R in the most POP contaminated snow petrels may be the consequence of an excess of ACTH input to adrenals. This suggests that POPs may alter the functioning of the pituitary by stimulating ACTH release and/or that POPs may mimic ACTH and bind to ACTH-R, which in that case would mobilize more ACTH-R from the adrenals of the most contaminated individuals. However this study is correlational, we cannot confirm without experimental support that the observed relationship is not the consequence of other intrinsic or extrinsic factors. Yet, an exacerbated secretion of CORT in response to a stressful event often mirrors poor fitness related traits as lowered parental investment (Angelier et al., 2009a; Bókony et al., 2009; Goutte et al., 2011b; Lendvai et al., 2007) or an impacted survival (Blas et al., 2007; Goutte et al., 2010; Romero, 2012). Nevertheless we did not find any relationship between POPs and parenting in terms of PRL concentration contrary to the study of Verreault et al. (2008) or egg-neglect behavior.

4.3. Hg: concentrations and relationships with stress-induced PRL and egg neglecting

Hg concentrations in incubating snow-petrels were within the range of those measured in the blood of south polar skuas breeding in Adélie land (2.15 \pm 0.17 $\mu g \cdot g^{-1}$ dw, Goutte et al., 2014). In comparison with an Arctic breeding seabird, we also found comparable concentrations in incubating black-legged kittiwakes (average 1.6 \pm 0.5 $\mu g \cdot g^{-1}$ dw). Contrary to other studies on free-ranging birds (Franceschini et al., 2009; Herring et al., 2012; Wada et al., 2009), we did not find any relationship between Hg and CORT secretion. Hg is well-known for its negative effects on breeding (reviewed in Tan et al., 2009). However, to the best of our knowledge, no studies have described relationships between Hg and PRL in free-living organisms. In humans, urinary Hg concentration was negatively correlated to plasma PRL (De Burbure and Bernard, 2006; Lucchini et al., 2002, 2003). In the present study, we found a similar relationship in incubating male snow petrels: increasing Hg concentrations were related to decreasing stress-induced PRL concentrations.

PRL is an anterior pituitary hormone, and a previous study on polar seabirds has described relationships between Hg and another anterior pituitary hormone: luteinizing hormone (LH, Tartu et al., 2013). Hg seemed to disrupt LH secretion via a lack of Gonadotropin-Releasing-Hormone (GnRH) input from the hypothalamus (Tartu et al., 2013). GnRH release is controlled by an area of the hypothalamus called zona incerta (Ben-Jonathan and Hnasko, 2001). Interestingly, this area also participates in the secretion of dopamine, a neuro-transmitter which is the principal antagonist of PRL (reviewed in Ben-Jonathan and Hnasko, 2001). Moreover, it has been well established that organic and inorganic Hg can stimulate the spontaneous release of dopamine in laboratory rodents (Faro et al., 1997, 2000, 2007; Minnema et al., 1989) but also in wild larvae of a fish (Fundulus heteroclitus, Zhou et al., 1999) and in wild American minks Mustela vison, where Hg induced a decrease of dopaminergic receptors and ligand affinity interpreted as an adaptive mechanism to prevent the hyperstimulation of the dopaminergic system (Basu et al., 2005). Additionally, when subjected to a stress, dopamine concentrations in blood increase (e.g. Finlay and Zigmond, 1997). Stress-induced dopamine synthesis in male snow petrels may thus be enhanced by Hg contamination, and result in a decrease of stress-induced PRL concentrations but not baseline or post-stress PRL concentrations. The fact that the most polluted birds quickly decrease their PRL concentrations when exposed to stress may highly affect their parental investment: they would be more likely to neglect their egg than less polluted birds. This goes together with the fact that in males, where PRL concentrations were lower than in females, the most polluted individuals were more likely to neglect their egg. In females, PRL concentrations and egg-neglect behavior were not related to Hg, maybe their PRL concentrations remained sufficiently high to prevent egg-neglect, a behavior associated with poor hatching success and chick mortality (Boersma and Wheelwright, 1979; Angelier et al., 2007).

5. Conclusion

In conclusion, there were no relationships between age and POPs or Hg, which is in line with most other studies. However we report significant relationships between contaminants and hormones involved in reproductive decisions. Over time, the action of POPs and Hg may jeopardize the maintenance of long-lived species populations. Indeed in long-lived species, that are expected to maximize their own survival rather than that of their brood, an exacerbated stress response as a consequence of POPs contamination and a decrease of PRL for the most Hg polluted males, are additional threats that may encourage individuals to refrain from breeding or desert their brood. To confirm the reported relationships, this study would greatly benefit from further experimental support.

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Conflict of interest statement

The authors declare no conflict of interest.

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References

Angelier F, Chastel O. Stress, prolactin and parental investment in birds: a review. Gen Comp Endocrinol 2009;163:142–8. http://dx.doi.org/10.1016/j.ygcen.2009.03.028.

Angelier F, Wingfield JC. Importance of the glucocorticoid stress response in a changing world: theory, hypotheses and perspectives. General and Comparative Endocrinology, 10th International Symposium on Avian. Endocrinology 2013;190:118–28.

Angelier F, Moe B, Weimerskirch H, Chastel O. Age-specific reproductive success in a long-lived bird: do older parents resist stress better? J Anim Ecol 2007;76:1181–91. http://dx.doi.org/10.1111/j.1365-2656.2007.01295.x.

Angelier F, Clément-Chastel C, Welcker J, Gabrielsen GW, Chastel O. How does corticosterone affect parental behaviour and reproductive success? A study of prolactin in black-legged kittiwakes. Funct Ecol 2009a;23:784–93. http://dx.doi.org/10.1111/j.1365-2435.2009.01545.x.

Angelier F, Moe B, Blanc S, Chastel O. What factors drive prolactin and corticosterone responses to stress in a long-lived bird species (snow petrel *Pagodroma nivea*)? Physiol Biochem Zool 2009b;82:590–602. http://dx.doi.org/10.1086/592846.

Angelier F, Wingfield JC, Weimerskirch H, Chastel O. Hormonal correlates of individual quality in a long-lived bird: a test of the "corticosterone–fitness hypothesis". Biol Lett 2010;6:846–9. http://dx.doi.org/10.1098/rsbl.2010.0376.

Angelier F, Wingfield JC, Trouvé C, de Grissac S, Chastel O. Modulation of the prolactin and the corticosterone stress responses: do they tell the same story in a long-lived bird, the Cape petrel? Gen Comp Endocrinol 2013;182:7–15. http://dx.doi.org/10.1016/j.ygcen.2012.10.008.

Barber JL, Sweetman AJ, van Wijk D, Jones KC. Hexachlorobenzene in the global environment: emissions, levels, distribution, trends and processes. Sci Total Environ 2005; 349:1–44. http://dx.doi.org/10.1016/j.scitotenv.2005.03.014.

- Barbraud C, Weimerskirch H. Contrasting effects of the extent of sea-ice on the breeding performance of an Antarctic top predator, the Snow Petrel *Pagodroma nivea*. J Avian Biol 2001;32:297–302. http://dx.doi.org/10.1111/j.0908-8857.2001.320402.x.
- Bargagli R. Environmental contamination in Antarctic ecosystems. Sci Total Environ 2008; 400:212–26. http://dx.doi.org/10.1016/j.scitotenv.2008.06.062.
- Basu N, Klenavic K, Gamberg M, O'Brien M, Evans D, Scheuhammer AM, et al. Effects of mercury on neurochemical receptor-binding characteristics in wild mink. Environ Toxicol Chem 2005;24:1444–50. http://dx.doi.org/10.1897/04-048R.1.
- Bearhop S, Ruxton GD, Furness RW. Dynamics of mercury in blood and feathers of great skuas. Environ Toxicol Chem 2000;19:1638–43. http://dx.doi.org/10.1002/etc. 5620190622.
- Ben-Jonathan N, Hnasko R. Dopamine as a prolactin (PRL) inhibitor. Endocr Rev 2001;22: 724–63. http://dx.doi.org/10.1210/er.22.6.724.
- Bergman Å, Heindel JJ, Jobling S, Kidd KA, Zoeller RT. State of the science of endocrine disrupting chemicals 2012: an assessment of the state of the science of endocrine disruptors prepared by a group of experts for the United Nations Environment Programme and World Health Organization; 2013 [xxv + 260 pp.].
- Beuschlein F, Fassnacht M, Klink A, Allolio B, Reincke M. ACTH-receptor expression, regulation and role in adrenocortical tumor formation. Eur J Endocrinol 2001;144: 199–206. http://dx.doi.org/10.1530/eje.0.1440199.
- Blas J, Bortolotti GR, Tella JL, Baos R, Marchant TA. Stress response during development predicts fitness in a wild, long lived vertebrate. Proc Natl Acad Sci U S A 2007;104: 8880–4. http://dx.doi.org/10.1073/pnas.0700232104.
- 8080–4. http://dx.doi.org/10.1073/phias.0700252104.
 Boersma PD, Wheelwright NT. Egg neglect in the Procellariiformes: reproductive adaptations in the Fork-tailed Storm-Petrel. Condor 1979:157–165.
- Bókony V, Lendvai ÁZ, Liker A, Angelier F, Wingfield JC, Chastel O. Stress response and the value of reproduction: are birds prudent parents? Am Nat 2009;173:589–98. http://dx.doi.org/10.1086/593131.
- Bonier F, Martin PR, Moore IT, Wingfield JC. Do baseline glucocorticoids predict fitness? Trends Ecol Evol 2009;24:634–42. http://dx.doi.org/10.1016/j.tree.2009.04.013.
- Borgå K, Wolkers H, Skaare JU, Hop H, Muir DCC, Gabrielsen GW. Bioaccumulation of PCBs in Arctic seabirds: influence of dietary exposure and congener biotransformation. Environ Pollut 2005;134:397–409. http://dx.doi.org/10.1016/j.envpol.2004.09.016.
- Breuner CW, Patterson SH, Hahn TP. In search of relationships between the acute adrenocortical response and fitness. Gen Comp Endocrinol 2008;157:288–95. http://dx.doi.org/10.1016/j.ygcen.2008.05.017.
- Buntin JD. Parental care: evolution, mechanisms, and adaptive significance. Academic Press: 1996.
- Bustamante P, Lahaye V, Durnez C, Churlaud C, Caurant F. Total and organic Hg concentrations in cephalopods from the North Eastern Atlantic waters: influence of geographical origin and feeding ecology. Sci Total Environ 2006;368:585–96. http://dx.doi.org/10.1016/j.scitotenv.2006.01.038.
- Bustnes JÖ, Bakken V, Erikstad KE, Mehlum F, Skaare JU. Patterns of incubation and nestsite attentiveness in relation to organochlorine (PCB) contamination in glaucous gulls. J Appl Ecol 2001;38:791–801. http://dx.doi.org/10.1046/j.1365-2664.2001.00633.x.
- Bustnes JO, Bakken V, Skaare JU, Erikstad KE. Age and accumulation of persistent organochlorines: a study of Arctic-breeding glaucous gulls (*Larus hyperboreus*). Environ Toxicol Chem 2003;22:2173–9. http://dx.doi.org/10.1897/02-456.
- Bustnes JO, Tveraa T, Henden JA, Varpe Ø, Janssen K, Skaare JU. Organochlorines in Antarctic and Arctic avian top predators: a comparison between the south polar skua and two species of Northern Hemisphere gulls. Environ Sci Technol 2006;40:2826–31. http://dx.doi.org/10.1021/es051920q.
- Bustnes JO, Tveraa T, Varpe Ø, Henden JA, Skaare JU. Reproductive performance and organochlorine pollutants in an Antarctic marine top predator: the south polar skua. Environ Int 2007;33:911–8. http://dx.doi.org/10.1016/j.envint.2007.04.010.
- Calamari D, Bacci E, Focardi S, Gaggi C, Morosini M, Vighi M. Role of plant biomass in the global environmental partitioning of chlorinated hydrocarbons. Environ Sci Technol 1991;25:1489–95. http://dx.doi.org/10.1021/es00020a020.
- Chastel O, Weimerskirch H, Jouventin P. High annual variability in reproductive success and survival of an Antarctic seabird, the snow petrel *Pagodroma nivea*. Oecologia 1993;94:278–85. http://dx.doi.org/10.1007/BF00341328.

 Chastel O, Lacroix A, Weimerskirch H, Gabrielsen GW. Modulation of prolactin but not
- Chastel O, Lacroix A, Weimerskirch H, Gabrielsen GW. Modulation of prolactin but not corticosterone responses to stress in relation to parental effort in a long-lived bird. Horm Behav 2005;47:459–66. http://dx.doi.org/10.1016/j.yhbeh.2004.10.009.
- Chaurand T, Weimerskirch H. Incubation routine, body mass regulation and egg neglect in the blue petrel *Halobaena caerulea*. Ibis 1994;136:285–90.
- Cherel Y, Mauget R, Lacroix A, Gilles J. Seasonal and fasting-related changes in circulating gonadal steroids and prolactin in king penguins, *Aptenodytes patagonicus*. Physiol Zool 1994;67:1154–73.
- Choi S-D, Baek S-Y, Chang Y-S, Wania F, Ikonomou MG, Yoon Y-J, et al. Passive air sampling of polychlorinated biphenyls and organochlorine pesticides at the Korean Arctic and Antarctic research stations: implications for long-range transport and local pollution. Environ Sci Technol 2008;42:7125–31. http://dx.doi.org/10.1021/es801004p.
- Clarke A, Harris CM. Polar marine ecosystems: major threats and future change. Environ Conserv 2003;30:1–25. http://dx.doi.org/10.1017/S0376892903000018.
- Corsolini S, Borghesi N, Ademolio N, Focardi S. Chlorinated biphenyls and pesticides in migrating and resident seabirds from East and West Antarctica. Environ Int 2011;37: 1329–35.
- Criscuolo F, Chastel O, Bertile F, Gabrielsen GW, Maho YL, Raclot T. Corticosterone alone does not trigger a short term behavioural shift in incubating female common eiders *Somateria mollissima*, but does modify long term reproductive success. J Avian Biol 2005;36:306–12. http://dx.doi.org/10.1111/j.0908-8857.2005.03371.x.
- De Burbure C, Bernard A. Prolactin changes as a consequence of chemical exposure: de Burbure and Bernard respond. Environ Health Perspect 2006;114:A574.

- Donaldson GM, Braune BM, Gaston AJ, Noble DG. Organochlorine and heavy metal residues in breast muscle of known-age thick-milled murres (*Uria lomvia*) from the Canadian Arctic. Arch Environ Contam Toxicol 1997;33:430–5. http://dx.doi.org/10.1007/s002449900273.
- Drent RH, Daan S. The prudent parent: energetic adjustments in avian breeding. Ardea 1980:68(1–4):225–52.
- Faro LRF, Durán R, do Nascimento JLM, Alfonso M, Picanço-Diniz CW. Effects of methyl mercury on the in vivo release of dopamine and its acidic metabolites DOPAC and HVA from striatum of rats. Ecotoxicol Environ Saf 1997;38:95–8. https://dx.doi.org/10.1006/eesa.1997.1567.
- Faro LRF, do Nascimento JLM, José JMS, Alfonso M, Durán R. Intrastriatal administration of methylmercury increases in vivo dopamine release. Neurochem Res 2000;25:225–9. http://dx.doi.org/10.1023/A:1007571403413.
- Faro LRF, Rodrigues KJA, Santana MB, Vidal L, Alfonso M, Durán R. Comparative effects of organic and inorganic mercury on in vivo dopamine release in freely moving rats. Braz J Med Biol Res 2007;40:1361–5. http://dx.doi.org/10.1590/S0100-879X2006005000157.
- Finlay JM, Zigmond MJ. The effects of stress on central dopaminergic neurons: possible clinical implications. Neurochem Res 1997;22:1387–94. http://dx.doi.org/10.1023/A:1022075324164.
- Franceschini MD, Lane OP, Evers DC, Reed JM, Hoskins B, Romero LM. The corticosterone stress response and mercury contamination in free-living tree swallows, *Tachycineta bicolor*. Ecotoxicology 2009;18:514–21. http://dx.doi.org/10.1007/s10646-009-0309-2.
- Frings CS, Fendley TW, Dunn RT, Queen CA. Improved determination of total serum lipids by the sulfo-phospho-vanillin reaction. Clin Chem 1972;18:673–4.
- Furness R, Hutton M. Pollutant levels in the great skua *Catharacta skua*. Environ Pollut 1979;19:261–8. http://dx.doi.org/10.1016/0013-9327(79)90118-6. Gabrielsen GW. Levels and effects of persistent organic pollutants in arctic animals. In:
- Gabrielsen GW. Levels and effects of persistent organic pollutants in arctic animals. In: Ørbæk DJB, Kallenborn DR, Tombre DI, Hegseth DEN, Falk-Petersen DS, Hoel DAH, editors. Arctic Alpine ecosystems and people in a changing environment. Springer Berlin Heidelberg; 2007. p. 377–412.
- Gonzáles-Solís J, Sanpera C, Ruiz X. Metals and selenium as bioindicators of geographic and trophic segregation in giant petrels *Macronectes* spp. Mar Ecol Prog Ser 2002; 244:257–64. http://dx.doi.org/10.3354/meps244257.
- Gordeev V. Pollution of the Arctic. Reg Environ Chang 2002;3:88–98. http://dx.doi.org/10.1007/s10113-002-0041-4.
- Goutte A, Angelier F, Welcker J, Moe B, Clément-Chastel C, Gabrielsen GW, et al. Longterm survival effect of corticosterone manipulation in Black-legged kittiwakes. Gen Comp Endocrinol 2010;167:246–51. http://dx.doi.org/10.1016/j.ygcen.2010.03.018.
- Goutte A, Kriloff M, Weimerskirch H, Chastel O. Why do some adult birds skip breeding? A hormonal investigation in a long-lived bird. Biol Lett 2011a;7:790–2. http://dx.doi.org/10.1098/rsbl.2011.0196.
- Goutte A, Antoine É, Chastel O. Experimentally delayed hatching triggers a magnified stress response in a long-lived bird. Horm Behav 2011b;59:167–73. http://dx.doi.org/10.1016/j.yhbeh.2010.11.004.
- Goutte A, Clément-Chastel C, Moe B, Bech C, Gabrielsen GW, Chastel O. Experimentally reduced corticosterone release promotes early breeding in black-legged kittiwakes. J Exp Biol 2011c;214:2005–13. http://dx.doi.org/10.1242/jeb.051979.
- Goutte A, Chevreuil M, Alliot F, Chastel O, Cherel Y, Eléaume M, et al. Persistent organic pollutants in benthic and pelagic organisms off Adélie Land, Antarctica. Mar Pollut Bull 2013;77:82–9.
- Goutte A, Bustamante P, Barbraud C, Delord K, Weimerskirch H, Chastel O. Demographic responses to mercury exposure in two closely-related Antarctic top predators. Ecology 2014. http://dx.doi.org/10.1890/13-1229.1.
- Groscolas R, Lacroix A, Robin J-P. Spontaneous egg or chick abandonment in energydepleted king penguins: a role for corticosterone and prolactin? Horm Behav 2008; 53:51–60. http://dx.doi.org/10.1016/j.yhbeh.2007.08.010.
- Guillette LJ, Gunderson MP. Alterations in development of reproductive and endocrine systems of wildlife populations exposed to endocrine-disrupting contaminants. Reproduction 2001;122:857–64. http://dx.doi.org/10.1530/rep.0.1220857.
- Heidinger BJ, Chastel O, Nisbet ICT, Ketterson ED. Mellowing with age: older parents are less responsive to a stressor in a long-lived seabird. Funct Ecol 2010;24:1037–44. http://dx.doi.org/10.1111/j.1365-2435.2010.01733.x.
- Henny CJ, Hill EF, Hoffman DJ, Spalding MG, Grove RA. Nineteenth century mercury: hazard to wading birds and cormorants of the Carson River, Nevada. Ecotoxicology 2002; 11:213–31. http://dx.doi.org/10.1023/A:1016327602656.
- Henriksen EO, Gabrielsen GW, Skaare JU. Levels and congener pattern of polychlorinated biphenyls in kittiwakes (*Rissa tridactyla*), in relation to mobilization of body-lipids associated with reproduction. Environ Pollut 1996;92:27–37. http://dx.doi.org/10.1016/0269-7491(95)00087-9.
- Henriksen EO, Gabrielsen GW, Utne Skaare J. Validation of the use of blood samples to assess tissue concentrations of organochlorines in glaucous gulls, *Larus hyperboreus*. Chemosphere 1998;37:2627-43. http://dx.doi.org/10.1016/S0045-6535(98)00162-3.
- Herring G, Ackerman JT, Herzog MP. Mercury exposure may suppress baseline corticosterone levels in juvenile birds. Environ Sci Technol 2012;46:6339–46. http://dx.doi.org/10.1021/es300668c.
- Hutton M. Accumulation of heavy metals and selenium in three seabird species from the United Kingdom. Environ Pollut Ser A Ecol Biol 1981;26:129–45. http://dx.doi.org/10.1016/0143-1471(81)90043-X.
- Jenssen BM. Endocrine-disrupting chemicals and climate change: a worst-case combination for Arctic marine mammals and seabirds? Environ Health Perspect 2005;114: 76–80. http://dx.doi.org/10.1289/ehp.8057.
- Keller JM, Kucklick JR, Harms CA, McClellan-Green PD. Organochlorine contaminants in sea turtles: correlations between whole blood and fat. Environ Toxicol Chem 2004; 23:726–38. http://dx.doi.org/10.1897/03-254.

- Kitaysky AS, Wingfield JC, Piatt JF. Dynamics of food availability, body condition and physiological stress response in breeding black-legged kittiwakes. Funct Ecol 1999;13: 577–84. http://dx.doi.org/10.1046/j.1365-2435.1999.00352.x.
- Koch KA, Wingfield JC, Buntin JD. Prolactin-induced parental hyperphagia in ring doves: are glucocorticoids involved? Horm Behav 2004;46:498–505. http://dx.doi.org/10.1016/j.yhbeh.2004.02.012.
- Lendvai AZ, Giraudeau M, Chastel O. Reproduction and modulation of the stress response: an experimental test in the house sparrow. Proc R Soc B 2007;274:391–7. http://dx.doi.org/10.1098/rspb.2006.3735.
- Lormée H, Jouventin P, Trouve C, Chastel O. Sex-specific patterns in baseline corticosterone and body condition changes in breeding red-footed boobies *Sula sula*. Ibis 2003;145:212–9. http://dx.doi.org/10.1046/j.1474-919X.2003.00106.x.
- Love OP, Shutt LJ, Silfies JS, Bortolotti GR, Smits JEG, Bird DM. Effects of dietary PCB exposure on adrenocortical function in captive American kestrels (*Falco sparverius*). Ecotoxicology 2003:12:199–208. http://dx.doi.org/10.1023/A:1022502826800.
- Ecotoxicology 2003;12:199–208. http://dx.doi.org/10.1023/A:1022502826800. Lucchini R, Cortesi I, Facco P, Benedetti L, Camerino D, Carta P, et al. Neurotoxic effect of exposure to low doses of mercury. Med Lav 2002;93:202–14.
- Lucchini R, Calza S, Camerino D, Carta P, Decarli A, Parrinello G, et al. Application of a latent variable model for a multicenter study on early effects due to mercury exposure. Neurotoxicology 2003;24:605–16. http://dx.doi.org/10.1016/S0161-813X(03) 00048-2.
- Minnema DJ, Cooper GP, Greenland RD. Effects of methylmercury on neurotransmitter release from rat brain synaptosomes. Toxicol Appl Pharmacol 1989;99:510–21. http:// dx.doi.org/10.1016/0041-008X(89)90158-0.
- Moline MA, Karnovsky NJ, Brown Z, Divoky GJ, Frazer TK, Jacoby CA, et al. High latitude changes in ice dynamics and their impact on polar marine ecosystems. Ann N Y Acad Sci 2008;1134:267–319. http://dx.doi.org/10.1196/annals.1439.010.
- Newton I, Bogan J, Marquiss M. Organochlorine contamination and age in sparrowhawks. Environ Pollut Ser A Ecol Biol 1981;25:155–60. http://dx.doi.org/10.1016/0143-1471(81)90016-7.
- Noël M, Barrett-Lennard L, Guinet C, Dangerfield N, Ross PS. Persistent organic pollutants (POPs) in killer whales (Orcinus orca) from the Crozet Archipelago, southern Indian Ocean. Mar Environ Res 2009;68:196–202. https://dx.doi.org/10.1016/j.marenvres.2009.06.009.
- Nordstad T, Moe B, Bustnes JO, Bech C, Chastel O, Goutte A, et al. Relationships between POPs and baseline corticosterone levels in black-legged kittiwakes (*Rissa tridactyla*) across their breeding cycle. Environ Pollut 2012;164:219–26. http://dx.doi.org/10.1016/j.envpol.2012.01.044.
- Odermatt A, Gumy C. Glucocorticoid and mineralocorticoid action: why should we consider influences by environmental chemicals? Biochem Pharmacol 2008;76: 1184–93. http://dx.doi.org/10.1016/j.bcp.2008.07.019.
- Ottinger MA, Abdelnabi M, Quinn M, Golden N, Wu J, Thompson N. Reproductive consequences of EDCs in birds: what do laboratory effects mean in field species? Neurotoxicol Teratol 2002;24:17–28. http://dx.doi.org/10.1016/S0892-0362(01) 00195-7
- Ottinger MA, Carro T, Bohannon M, Baltos L, Marcell AM, McKernan M, et al. Assessing effects of environmental chemicals on neuroendocrine systems: potential mechanisms and functional outcomes. Gen Comp Endocrinol 2013;190:194–202. http://dx.doi.org/10.1016/j.ygcen.2013.06.004.
- Pauwels A, Covaci A, Weyler J, Delbeke L, Dhont M, Sutter PD, et al. Comparison of persistent organic pollutant residues in serum and adipose tissue in a female population in Belgium, 1996–1998. Arch Environ Contam Toxicol 2000;39:265–70. http://dx.doi.org/10.1007/s002440010104.
- Penhoat A, Jaillard C, Saez JM. Corticotropin positively regulates its own receptors and cAMP response in cultured bovine adrenal cells. Proc Natl Acad Sci U S A 1989;86: 4978–81.
- Ricklefs RE, Wikelski M. The physiology/life-history nexus. Trends Ecol Evol 2002;17: 462–8. http://dx.doi.org/10.1016/S0169-5347(02)02578-8.
- Risebrough RW, De Lappe BW, Schmidt TT. Bioaccumulation factors of chlorinated hydrocarbons between mussels and seawater. Mar Pollut Bull 1976;7:225–8. http://dx.doi.org/10.1016/0025-326X(76)90266-6.
- Romero LM. Using the reactive scope model to understand why stress physiology predicts survival during starvation in Galápagos marine iguanas. Gen Comp Endocrinol 2012; 176:296–9. http://dx.doi.org/10.1016/j.ygcen.2011.11.004.
- Romero LM, Reed JM. Collecting baseline corticosterone samples in the field: is under 3 min good enough? Comp Biochem Physiol A Mol Integr Physiol 2005;140:73–9. http://dx.doi.org/10.1016/j.cbpb.2004.11.004.
- Rowe CL. "The calamity of so long life": life histories, contaminants, and potential emerging threats to long-lived vertebrates. Bioscience 2008;58:623–31. http://dx.doi.org/
- Ryan PG, Watkins BP. Snow petrel breeding biology at an inland site in continental Antarctica. Colon Waterbirds 1989;12:176. http://dx.doi.org/10.2307/1521338.
- Smetacek V, Nicol S. Polar ocean ecosystems in a changing world. Nature 2005;437: 362–8. http://dx.doi.org/10.1038/nature04161.
- Solheim SA. The preen gland an organ for excretion of persistent organic pollutants in black-legged kittiwake (*Rissa tridactyla*). Master thesis University of Bergen; 2010.
- Szumiło E, Szubska M, Meissner W, Bełdowska M, Falkowska L. Mercury in immature and adults herring gulls (*Larus argentatus*) wintering on the Gulf of Gdańsk area. Ocean Hydro 2013;42:260–7. http://dx.doi.org/10.2478/s13545-013-0082-y.

- Tan SW, Meiller JC, Mahaffey KR. The endocrine effects of mercury in humans and wildlife. Crit Rev Toxicol 2009;39:228–69. http://dx.doi.org/10.1080/10408440802233259.
- Tapie N, Menach KL, Pasquaud S, Elie P, Devier MH, Budzinski H. PBDE and PCB contamination of eels from the Gironde estuary: from glass eels to silver eels. Chemosphere 2011;83:175–85. http://dx.doi.org/10.1016/j.chemosphere.2010.12.044.
- Tartu S, Goutte A, Bustamante P, Angelier F, Moe B, Clément-Chastel C, et al. To breed or not to breed: endocrine response to mercury contamination by an Arctic seabird. Biol Lett 2013;9:20130317. http://dx.doi.org/10.1098/rsbl.2013.0317.
- Tartu S, Angelier F, Herzke D, Moe B, Bech C, Gabrielsen GW, et al. The stress of being contaminated? Adrenocortical function and reproduction in relation to persistent organic pollutants in female black legged kittiwakes. Sci Total Environ 2014;476–477: 553–60. http://dx.doi.org/10.1016/j.scitotenv.2014.01.060.
- Tavares S, Xavier JC, Phillips RA, Pereira ME, Pardal MA. Influence of age, sex and breeding status on mercury accumulation patterns in the wandering albatross *Diomedea exulans*. Environ Pollut 2013;181:315–20. http://dx.doi.org/10.1016/j.envpol.2013.06.032
- Thompson DR, Hamer KC, Furness RW. Mercury accumulation in great skuas *Catharacta* skua of known age and sex, and its effects upon breeding and survival. J Appl Ecol 1991: 28:672–84
- Tyler CR, Jobling S, Sumpter JP. Endocrine disruption in wildlife: a critical review of the evidence. Crit Rev Toxicol 1998;28:319–61. http://dx.doi.org/10.1080/10408449891344236.
- Van de Merwe JP, Hodge M, Olszowy HA, Whittier JM, Lee SY. Using blood samples to estimate persistent organic pollutants and metals in green sea turtles (*Chelonia mydas*). Mar Pollut Bull 2010:60:579–88.
- Van den Brink NW. Directed transport of volatile organochlorine pollutants to Polar Regions: the effect on the contamination pattern of Antarctic seabirds. Sci Total Environ 1997;198:43–50. http://dx.doi.org/10.1016/S0048-9697(97)05440-5.
- Van den Brink NW, van Franeker JA, de Ruiter-Dijkman EM. Fluctuating concentrations of organochlorine pollutants during a breeding season in two Antarctic seabirds: Adelie penguin and southern fulmar. Environ Toxicol Chem 1998;17:702–9. http://dx.doi.org/10.1002/etc.5620170426.
- Verboven N, Verreault J, Letcher RJ, Gabrielsen GW, Evans NP. Nest temperature and parental behaviour of Arctic-breeding glaucous gulls exposed to persistent organic pollutants. Anim Behav 2009;77:411–8. http://dx.doi.org/10.1016/j.anbehav.2008.10.022
- Verboven N, Verreault J, Letcher RJ, Gabrielsen GW, Evans NP. Adrenocortical function of Arctic-breeding glaucous gulls in relation to persistent organic pollutants. Gen Comp Endocrinol 2010;166:25–32. http://dx.doi.org/10.1016/j.ygcen.2009.11.013.
- Verreault J, Letcher RJ, Muir DCG, Chu S, Gebbink WA, Gabrielsen GW. New organochlorine contaminants and metabolites in plasma and eggs of glaucous gulls (*Larus hyperboreus*) from the Norwegian Arctic. Environ Toxicol Chem 2005;24:2486. http://dx.doi.org/10.1897/05-067R.1.
- Verreault J, Verboven N, Gabrielsen GW, Letcher RJ, Chastel O. Changes in prolactin in a highly organohalogen contaminated Arctic top predator seabird, the glaucous gull. Gen Comp Endocrinol 2008;156:569–76. http://dx.doi.org/10.1016/j.ygcen.2008.02. 013
- Wada H, Cristol DA, McNabb FMA, Hopkins WA. Suppressed adrenocortical responses and thyroid hormone levels in birds near a mercury-contaminated river. Environ Sci Technol 2009;43:6031–8. http://dx.doi.org/10.1021/es803707f.
- Walker CH, Sibly RM, Hopkin SP, Peakall DB. Principles of ecotoxicology. 4th ed. CRC Press: 2012.
- Wania F. Assessing the potential of persistent organic chemicals for long-range transport and accumulation in Polar Regions. Environ Sci Technol 2003;37:1344–51. http://dx.doi.org/10.1021/es026019e.
- Wania F, MacKay D. Peer reviewed: tracking the distribution of persistent organic pollutants. Environ Sci Technol 1996;30:390A–6A. http://dx.doi.org/10.1021/es962399q.
- Wayland M, Garcia-Fernandez AJ, Neugebauer E, Gilchrist HG. Concentrations of cadmium, mercury and selenium in blood, liver and kidney of common eider ducks from the Canadian Arctic. Environ Monit Assess 2001;71:255–67. http://dx.doi.org/10. 1023/A:1011850000360.
- Weimerskirch H, Lallemand J, Martin J. Population sex ratio variation in a monogamous long-lived bird, the wandering albatross. J Anim Ecol 2005;74:285–91. http://dx.doi.org/10.1111/j.1365-2656.2005.00922.x.
- Wingfield J. Modulation of the adrenocortical response to stress in birds. Perspectives in comparative endocrinology; 1994. p. 520–8.
- Wingfield JC, Sapolsky RM. Reproduction and resistance to stress: when and how. J Neuroendocrinol 2003;15:711–24. http://dx.doi.org/10.1046/j.1365-2826.2003. 01033.x.
- Xie Z, Zhang P, Sun L, Xu S, Huang Y, He W. Microanalysis of metals in barbs of a snow petrel (*Pagodroma nivea*) from the Antarctica using synchrotron radiation X-ray fluorescence. Mar Pollut Bull 2008;56:516–24. http://dx.doi.org/10.1016/j.marpolbul.2007.11.015.
- Zhou T, Rademacher DJ, Steinpreis RE, Weis JS. Neurotransmitter levels in two populations of larval Fundulus heteroclitus after methylmercury exposure. Comp Biochem Physiol C Pharmacol Toxicol Endocrinol 1999(124):287–94. http://dx.doi.org/10.1016/S0742-8413(99)00077-8.