Trace Elements in Three Marine Birds Breeding on Reunion Island (Western Indian Ocean): Part 2—Factors Influencing Their Detoxification

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Abstract. Seabird tissues collected between 2002 and 2004 from Barau’s Petrel (Pterodroma baraui), Audubon’s Shearwater (Puffinus lherminieri bailloni), and White-Tailed Tropicbird (Phaethon lepturus) colonies on Reunion Island were analyzed for metallothioneins (MTs) and trace element content. The subcellular distribution between soluble and insoluble fractions of Cd, Cu, Fe, Mn, Se, and Zn was determined in liver and kidney. In both, the soluble fraction of the cell concentrated most of the Cd and Se, whereas Fe, Mn, and Zn were preferentially accumulated in the insoluble fraction. The distribution of these elements varied with the tissue, age of the bird, and species. Furthermore, the distributions of Fe and Mn were somewhat influenced by the bird’s physical condition. MT levels were measured in the soluble fraction after heat denaturation. The levels of these proteins varied from 5.5 ± 2.7 mg.g⁻¹ dry weight (dw) to 11.4 ± 6.2 mg.g⁻¹ dw depending on the species and the tissue considered. MT levels were significantly different between liver and kidney only in the White-Tailed Tropicbird. In the three species, MT levels in kidney were significantly higher in adult than juvenile birds. The bird’s weight also had an influence on hepatic and renal MT levels, but not the sex nor the reproductive status. The implication of MTs in Cu and Zn homeostasis and Cd and Hg detoxification are discussed. In addition, clues on Hg regulation by Se were found, especially in Barau’s Petrel, where the levels of these two elements were significantly correlated.

Marine birds accumulate high levels of trace elements because of their position in marine food webs and their long life span. Cd and Hg, which are naturally present in the environment, are toxic to marine biota. Essential elements, such as Cu, Fe, Mn, Se, and Zn are necessary for the metabolism but can cause adverse effects when their concentrations in an organism become excessive. Quantification of trace element levels in different tissues of an organism is an indicator of the bioavailable fraction of the element in the environment (Hogstrand and Haux 1990). Nevertheless, these measures do not indicate whether an organism is stressed or affected by the contaminant. The physicochemical form of storage of the element may however give information on the tissue(s) where detoxification takes place.

In marine vertebrates, detoxification is the result of many processes. One of the most commonly studied is the binding of metals to metallothioneins (MTs) (Mason and Jenkins 1995). Metallothioneins constitute a class of soluble low–molecular weight metalloproteins characterized by their heat stability, high cystein content, and lack of aromatic amino acid (Hamer 1986). These proteins play an important role in the transport and storage of trace elements. They also provide protection against the toxic effects of certain metals by sequestrating and decreasing the amount of free metal ions and acting as a “rescue” function for structures impaired by inappropriate metal binding (Hamilton and Mehrle 1986; Vallee 1995; Rosésjádi 1996). Since their discovery in 1957, the existence of MTs has been firmly established in a large number of animals (Hamza-Chaffai et al. 1995). MTs has been shown to occur in liver and/or kidneys of various species of seabirds, such as Leach’s Storm Petrel (Oceanodroma leucorhoa), the Atlantic Puffin (Fratercula arctica), the Double Crested Cormorant (Phalacrocorax auritus), the Herring Gull (Larus argentatus), the Black-Headed Gull (Larus ridibundus), the Northern Fulmar (Fulmarus glacialis), the Common Guillemot (Uria aalge), the Kittiwake (Rissa tridactyla), and the Common Scoter (Melanitta nigra) (Osborn 1978; Elliott et al. 1992; Wenzel and Adelung 1996). Measurements of MT levels and their relationships with element levels constitute an attractive insight to trace element behavior in an organism.

Another common detoxification process relies on the insolubilization of metals as mineral concretions. A typical example is the formation of insoluble tiemannite granules after the binding of Hg by Se in the liver of many marine mammals and certain birds (Koeman et al. 1973; Martoja and Berry 1980; Nigro et al. 2002, Decataldo et al. 2004, Ikemoto et al. 2004). Enriched Cd granules are also known to exist in White-Sided dolphin (Lagenorhynchus acutus) kidneys (Gallien et al. 2001).
The nature of the detoxification process adopted by an organism gives insight into its degree of exposure to toxic elements. For example, MTs tend to take part in the detoxification of elements to which exposure is low or sudden (Chan et al. 1993), whereas granules attest to exposure to higher levels during a longer period of time (Palmisano et al. 1995).

Our work focused on three seabird species breeding on Reunion Island that differ in their dietary ecology and in their contamination by elements as discussed in the first part of this study (Kojadinovic et al. 2007). Barau’s Petrel (Pterodroma baraud) is an oceanic bird that feeds almost exclusively on cephalopods. Juvenile birds accumulate particularly high levels of Cd and Hg, and adult birds mainly have high Fe, Hg, and Se levels. The White-Tailed Tropicbird (Phaethon lepturus) also forages over oceanic waters but closer to Reunion Island and is less specialized on squid. Compared with the other two species, high levels of Cu, Fe, Mn, and Zn were observed in juvenile and adult birds. Finally, Aububon’s Shearwater (Puffinus lherminieri bailloni) forages closest to the island, feeds as much on cephalopods as it does on fish (Bailey 1967; Jaquemet et al. 2004), and shows the lowest Hg levels (Kojadinovic et al. 2007). Information on species-specific differences in the sensitivity to trace elements would be important in evaluating the adverse effects of elements on wild birds.

Thus, in this second part of our work, we examined trace element regulation in Reunion seabirds by studying the subcellular distribution of trace element and MT levels in liver and kidney as well as factors influencing MT levels. The interaction between MTs and Cd, Cu, Hg, and Zn, and the relationships between elements, will be discussed to understand the potential detoxification strategies used by these birds.

Materials and Methods

Study Site and Species

Barau’s Petrels, Aububon’s Shearwaters, and White-Tailed Tropicbirds used in this study originated from colonies established on Reunion Island (21°7'S, 55°33'E) in the Western Indian Ocean. As noted earlier, these seabird species have contrasting dietary ecologies during their breeding season (approximately 3 months) during which they feed more or less in proximity to the island. Outside of their breeding period, individuals scatter in the waters of the Indian Ocean (Barré et al. 1996; Stahl and Bartle 1991). During that time, dietary habits of the three species remain unknown.

Bird Sampling

Fifty-two Barau’s Petrels, 61 Aububon’s Shearwaters, and 49 White-Tailed Tropicbirds were sampled (see Kojadinovic et al. 2007). Because these birds were found before or shortly after accidental death, we are confident that the conditions under which the birds died had no effect on their elemental and MT levels. Each bird was measured, weighed, and aged. Adults were sexed and their reproductive status noted. Liver, kidneys, and pectoral muscles were removed during dissection, weighed, and frozen before analysis. At this stage, muscular condition (MC) and body condition (BC) were evaluated by MC and BC indices, respectively (Kojadinovic et al. 2007).

Sample Preparation

To prepare for elemental and MT determination, livers and kidneys were blended, lyophilized, and ground to a fine powder. Aliquots of approximately 100 mg were homogenized on ice in 6 ml 100 mM Tris buffer with β-mercaptoethanol at pH = 8.1 and then centrifuged. Metal content was determined in the pellet (C1). The obtained supernatant (S1) was divided for metal (5 ml) and MT (1 ml) analyses. Before MT dosage, the supernatant aliquot was submitted to heat denaturation and centrifugated to separate the heat-stable proteins from the denatured proteins (Fig. 1). The supernatant stemming from the second centrifugation (S2) was frozen (−80°C) until MT quantification.

The analysis of Cd, Cu, Fe, Mn, Se, and Zn conducted in S1 and C1 called for an extra step in the preparation protocol. Both fractions were completely dried and digested with 1 ml 15 N nitric acid at 60°C for 48 hours before being diluted in 10 ml deionized water.

MTs were not analyzed in muscle tissue. Muscle samples were thus dried at 55°C in an oven before being ground. Aliquots of approximately 100 mg were digested with 3.5 ml 15 N nitric acid and diluted in 10 ml deionized water.

Accuracy and reproducibility of the preparation were tested by preparing 10 replicates of reference standards (National Research Council, Canada) and 11 blanks along with each set of samples. “The sample preparation was done in “metal-free” conditions (Kojadinovic et al. 2007).

Metal Analysis

Cd, Cu, Fe, Mn, Se, and Zn were analyzed in S1 and C1 by inductively coupled plasma atomic emission spectrometry (ICP-AES Varian Vista Pro CCD). Total levels were obtained by adding the elemental masses found in S1 and C1 and dividing by the aliquot mass. Total Hg analyzes were carried out with an advanced mercury analyzer (ALTEC AMA 254). Detection limits and recovery rates are given in the first part of this study (Kojadinovic et al. 2006).

Metallothionein Analysis

Of the 162 birds, 70 were analyzed for their hepatic and renal MT levels. Differential pulse polarographic analysis (DPP) was used to determine the amount of MTs in the heat-denatured soluble fraction (S2). DPP is a technique based on −SH compound determination according to Budička reaction (Budička 1933) as described by Thompson and Cosson (1984). Model 303A static mercury drop electrode was used. Certified rabbit liver MTs (Sigma Chemical Co., St. Louis, MO) were used to carry out the calibration according to the method of standard additions. The system consisted of a beveled capillary, a Hg working electrode, a platinum counter electrode, and an Ag/AgCl reference electrode. Results are expressed in mg of MTs per g of dry homogenized tissue. For comparison with concentrations given in relation to wet weight, refer to the mean moisture contents given hereafter: from 66% to 72% in liver, from 73% to 78% in kidney, and from 65% to 71% in muscle.

Statistical Analyses

Statistical analyses were performed using the GNU R statistical system (R Development Core Team 2005). Data were first checked for normality by means of Shapiro-Wilk test. In cases of nondeparture from normality, parametric tests were used in the subsequent
analyses. When Shapiro-Wilk test p values were < 0.05, nonparametric analogues were used. Before the use of analysis of variance (ANOVA) for independent samples, in addition to normality, the homogeneity of the variances of the tested samples was checked by means of Bartlett test. In case of departure from normality or non homogeneity of the variances, Kruskal-Wallis tests were applied instead.

The significance of differences of MT levels between liver and kidney was tested by means of student $t$-test or Wilcoxon (W) test according to normality. The influence of species and MC on MT content was tested by means of ANOVA or Kruskal-Wallis tests followed by Tukey's Honestly Significant Difference (HSD) test. The influence of age, sex, and reproductive status was tested by means of student $t$-test or W test. For each species, dependencies between trace element and MT levels were studied by means of Pearson's linear correlation coefficient. To study the factors influencing the distribution of element levels between soluble and insoluble fractions, the latter tests were applied to the ratio between the mass of an element in the soluble fraction and the total mass of this element in both fractions.

Levels of significance of the null hypotheses associated with these tests were divided into classes of $p$ values represented by the following codes: NS $\geq 0.05$; * $< 0.05$; ** $< 0.01$; *** $< 0.001$. SD stands for standard deviation, and CV stands for coefficient of variation.

Subcellular Elemental Distribution

The subcellular distribution of analyzed elements is given in Table 1 as the percentage of the element recovered in the soluble fraction versus the total amount of metal in the given tissue. It was assumed that, after centrifugation at 1800 g for 60 minutes, the nuclei and debris of various origins would sediment to constitute C1, whereas S1 corresponded to the soluble fraction that contained the MTs. Figs. 2 and 3 illustrate the levels of these elements in the soluble fraction in relation to the whole-tissue levels.

The distribution pattern of trace elements between both fractions was similar in the three species. Cd, Se, and hepatic Cu were more present in S1, whereas Fe, Mn, and renal Cu and Zn were in a larger part found in C1. In most cases, the proportion of trace elements in the soluble fraction was higher in liver than kidney. Furthermore, the concentrations of each element in S1 increased with the total cellular elemental concentrations (Figs. 2 and 3). Moreover, the linear correlations between levels in the soluble fraction and in the whole

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**Fig. 1.** Sample preparation for trace element and MT analysis in seabird tissues
Fig. 2. Element levels (µg g⁻¹ dw) in the soluble fraction in relation to their levels in the whole liver of Barau’s Petrel, Audubon’s Shearwater, and White-tailed Tropicbird.
Fig. 3. Element levels (μg.g⁻¹ dw) in the soluble fraction in relation to their levels in the whole kidney of Barau’s Petrel, Audubon’s Shearwater and White-tailed Tropicbird.
tissue were generally better in liver than kidney. They were particularly good for Cd, Cu, Se, and Zn (Figs. 2 and 3).

Despite the general similarity of the results in three species, some differences were observed between the petrel, the shearwater, and the tropicbird. The distribution of hepatic Cd and Cu was significantly different between Audubon’s Shearwater and the White-Tailed Tropicbird ($P_{\text{Tukey}}$: **); in kidney, the two Procellariiformes differed in the distribution of Cu ($P_{\text{Tukey}}$: *) and Fe ($P_{\text{Tukey}}$: *); the shearwater differed from the tropicbird in the distribution of Cu and Zn ($P_{\text{Tukey}}$: **) and Barau’s Petrel differed from the tropicbird in the distribution of Mn ($P_{\text{Tukey}}$: **).

The only significant influence of age on trace element distribution, in all species combined, was found for Fe ($P_{\text{Fisher}}$: *), Mn ($P_{\text{Fisher}}$: *), and Se ($P_{\text{Wilcoxon}}$: ***) in liver. The accumulation of these elements in the soluble fraction was higher in adult than in juvenile birds.

Sex, across all, did not have an impact on the distribution of trace elements, except for hepatic Zn ($P_{\text{Fisher}}$: **), which was found to accumulate significantly more in the soluble fraction in female than in male birds. In the same way, the reproductive status of adult birds did not influence the distribution of trace elements in the majority of cases. It was, however, observed that renal Mn and Zn levels were higher (respectively, $P_{\text{Wilcoxon}}$: ** and *) in the insoluble fraction of incubating birds.

The influence of MC on the distribution of Cd, Cu, Fe, Mn, Se, and Zn was tested. Hepatic Mn, and hepatic and renal Fe, were proportionally more present in the soluble fraction than the insoluble fraction in birds with good MC (respectively, $P_{\text{Kruskal-Wallis}}$: ***, $P_{\text{ANOVA}}$: *, and $P_{\text{ANOVA}}$: *). Although less obvious, the same trend was observed for Fe and Mn with BC. In the tropicbird, hepatic and renal Fe and liver Mn were preferentially accumulated in S1 in larger birds. Indeed, significant positive correlations were found between the bird’s body weight and hepatic levels of Fe in S1 ($r = 0.723$, *), the bird’s weight and kidney Fe ($r = 0.347$, *), and the bird’s weight and liver Mn ($r = 0.651$, *). The opposite tendency was noticed for kidney and liver Zn in Barau’s Petrel (respectively, $r = -0.971$, *, $r = 0.93$, *), but this may have resulted from the low number of samples.

### Metallothioneins

MT levels found in liver and kidney of Barau’s Petrels, Audubon’s Shearwaters, and White-Tailed Tropicbirds are listed in Table 2. The linear correlations between MT and element levels are listed in Table 3. Positive correlations between total Cd, Cu, Hg, and Zn levels and MT levels in liver and kidney indicate that MT content of these organs increases with metal levels. At the species level, the highest MT concentrations were found in Barau’s Petrel (Table 2), which exhibited the highest Hg levels of the three species (Kojadinovic et al. 2007).

MT levels were significantly different between liver and kidney in the White-Tailed Tropicbird ($P_{\text{Tukey}}$: **). However,
this was not the case for the two Procellariiform species. MT levels were well correlated between both tissues in the three bird species (Table 3).

In the three species combined, renal MT levels were significantly higher in adult, than in juvenile birds (PW: ***), whereas there were no significant differences in liver. As it was observed for trace element levels (Kojadinovic et al. 2006), neither the sex nor the reproductive status of the adult birds of each species had any influence on the MT levels.

BC did not seem to have any influence on MT levels of each species (Table 3). However, MC had an impact on MT concentrations in liver and kidney, with lower MT levels being found in birds with the best MC. Furthermore, significant negative correlations were found between the bird total weight and MT levels, especially in liver.

MT can bind atoms of Cd, Cu, Hg, and Zn (Hamer 1986). The number of atoms of Cd, Cu, and Zn was calculated per gram of dry tissue and summed (denoted by molCdCuHgZn in Table 3), to examine the potential linkage of MTs with the four atoms simultaneously. Although the degree of correlation varied from one species to the other, the same general pairs of well-correlated variables appeared, such as MT–Zn in liver and kidney. The summed Cd, Cu, Hg, and Zn atoms were also well correlated with MT levels in the corresponding tissue in all three species. Positive correlations were also found between MT levels in one tissue and metal levels in the other. For example, hepatic MT levels were correlated

<table>
<thead>
<tr>
<th>Barau’s Petrel (n = 15)</th>
<th>Audubon’s Shearwater (n = 20)</th>
<th>White-Tailed Tropicbird (n = 32)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Liver</strong></td>
<td><strong>Liver</strong></td>
<td><strong>Liver</strong></td>
</tr>
<tr>
<td>MT.L–Zn.L 0.852***</td>
<td>molCdCuHgZn.L - MT.L 0.772 ***</td>
<td>molCdCuHgZn.L - MT.L 0.947 ***</td>
</tr>
<tr>
<td>molCdCuHgZn.L - MT.L 0.835 ***</td>
<td>MT.L - Zn.L 0.715 ***</td>
<td>MT.L – Zn.L 0.924 ***</td>
</tr>
<tr>
<td>MT.L - Bird weight -0.726 **</td>
<td>MT.L - Cd.L 0.633 **</td>
<td>MT.L - Bird weight -0.74 ***</td>
</tr>
<tr>
<td>MT.L - Hg.L 0.696 **</td>
<td>MT.L - Bird weight -0.581 *</td>
<td>MT.L - Cu.L 0.667 ***</td>
</tr>
<tr>
<td>MT.L - Cd.L 0.622 *</td>
<td>MT.L - Hg.L 0.565 **</td>
<td>MT.L - BC -0.622 ***</td>
</tr>
<tr>
<td>MT.L - Cu.L -0.243 NS</td>
<td>MT.L - BC -0.554 **</td>
<td>MT.L - Cd.L 0.34 *</td>
</tr>
<tr>
<td>MT.L - BC -0.085 NS</td>
<td>MT.L - Cu.L 0.465 *</td>
<td>MT.L - Hg.L 0.339 *</td>
</tr>
<tr>
<td><strong>Kidney</strong></td>
<td><strong>Kidney</strong></td>
<td><strong>Kidney</strong></td>
</tr>
<tr>
<td>MT.K - Zn.K 0.802 ***</td>
<td>MT.K - Zn.K 0.803 ***</td>
<td>molCdCuHgZn.K - MT.K 0.866 ***</td>
</tr>
<tr>
<td>molCdCuHgZn.K - MT.K 0.74 ***</td>
<td>molCdCuHgZn.K - MT.K 0.776 ***</td>
<td>MT.K - Zn.K 0.834 ***</td>
</tr>
<tr>
<td>MT.K - Cd.K 0.541 *</td>
<td>MT.K - Hg.K 0.68 ***</td>
<td>MT.K - Cd.K 0.65 ***</td>
</tr>
<tr>
<td>MT.K - Bird weight -0.434 NS</td>
<td>MT.K - Cd.K 0.642 **</td>
<td>MT.K - Cu.K 0.623 ***</td>
</tr>
<tr>
<td>MT.K - BC 0.424 NS</td>
<td>MT.K - Cu.K 0.61 **</td>
<td>MT.K - Hg.K 0.546 ***</td>
</tr>
<tr>
<td>MT.K - Cu.K 0.3 NS</td>
<td>MT.K - Bird weight -0.506 *</td>
<td>MT.K - Bird weight -0.38 **</td>
</tr>
<tr>
<td>MT.K - Hg.K 0.25 NS</td>
<td>MT.K - BC -0.272 NS</td>
<td>MT.K - BC -0.328 NS</td>
</tr>
</tbody>
</table>

**Intertissue correlations**

| MT.L - Cd.M 0.74 ** | MT.L - Zn.M 0.866 *** | MT.L - Zn.M 0.872 *** |
| MT.K-Cd.M 0.718 ** | MT.L - Hg.M 0.774 *** | MT.K - Cd.M 0.8 *** |
| MT.K - MT.L 0.696 ** | MT.K - Hg.M 0.771 *** | MT.K-Cu.M 0.764 *** |
| ML.L - Cu.M 0.667 ** | MT.L - Cu.M 0.759 *** | MT.L - Zn.K 0.751 *** |
| MT.L - Zn.M 0.651 | MT.K - MT.L 0.747 *** | MT.L - Cu.M 0.69 *** |
| MT.L - Cu.M 0.645 ** | MT.L - Hg.K 0.687 *** | molCdCuHgZn.K - MT.L 0.665 *** |
| MT.K - Hg.L 0.632 | MT.L - Cu.K 0.655 ** | MT.K - Zn.M 0.653 *** |
| MT.L - Zn.K 0.631 | MT.L - Cd.M 0.648 *** | MT.K - MT.L 0.648 *** |
| molCdCuHgZn.K - MT.L 0.631 | molCdCuHgZn.K - MT.L 0.633 ** | MT.K - Cd.M 0.647 *** |
| MT.K - Zn.M 0.572 | MT.L - Zn.K 0.615 ** | MT.K - Hg.M 0.592 *** |
| MT.K - Hg.M 0.533 | MT.K - Zn.M 0.577 | MT.L - Hg.M 0.549 *** |
| molCdCuHgZnL - MT.K 0.5 | MT.K - Cu.M 0.558 | molCdCuHgZnL - MT.K 0.501 *** |
| MT.L - Cd.K 0.53 NS | MT.L - Cd.K 0.484 | MT.K – Cu.L 0.491 *** |
| MT.L - Hg.M 0.51 | MT.L - Cd.D 0.479 | MT.L - Cd.M 0.48 ** |
| MT.K - Cd.L 0.465 NS | MT.K - Zn.L 0.461 NS | MT.K - Zn.L 0.437 ** |
| MT.L - Cu.K 0.463 NS | MT.K - Hg.L 0.453 NS | MT.L - Hg.K 0.401 * |
| MT.L - Hg.K 0.443 NS | molCdCuHgZnL - MT.K 0.427 NS | MT.L - Cu.K 0.369 *** |
| MT.K - Zn.L 0.441 NS | MT.K - Cd.M 0.391 NS | MT.K - Hg.L 0.352 ** |
| MT.K - Cu.L -0.22 NS | MT.K - Cu.L 0.261 NS | MT.L - Cd.K 0.339 * |

* The letters “L”, “K” or “M” placed after the abbreviation of an element indicate the tissues (liver, kidney, and muscle, respectively) in which the element is considered.

NS p > 0.05.

* p < 0.05.

** p < 0.01.

*** p < 0.001.
to muscular Cu and Zn levels in all species, and renal MT levels were correlated to Cd levels in muscle, liver, and kidney.

Discussion

In addition to its concentration in a tissue, the toxicity of an element is subject to a number of factors such as the interaction of this element with others, its chemical speciation, and its availability, which is dictated by its location in a tissue as well as at a cellular level. The study of element subcellular distribution, along with the investigation of potential detoxification processes, is necessary to understanding the path followed by each element in an organism.

The most evident subcellular distribution trend was that, in all three species, the proportion of each element in the soluble fraction was higher in the liver than in the kidney (Table 1). The function of each organ probably explains this distribution pattern. The liver is an organ of great metabolic activity and a reservoir for essential elements that must be available and are thus located in the soluble fraction. The kidney is an organ of which the principal function is the excretion of the organism’s waste material, but it is also a “sink” for elements that cannot be excreted.

In the three species, Fe, Mn, and renal Cu were in large part found in the insoluble fraction (Table 1). The trends observed for these essential metals were similar to those observed in various seabirds and other marine animals (Anan et al. 2002; Ikemoto et al. 2004; Nam et al. 2005) and confirm that these elements are part of the composition of organelle and membrane debris that sedimented during centrifugation. Cu concretions may also account for the renal Cu found in C1 (Bremner and Mehra 1991). The MC index had an impact on the distribution of hepatic Mn as well as hepatic and renal Fe, which were proportionally more present in the soluble fraction than in insoluble fraction in birds with good MC. Given the importance of pectoral muscles in flight, it can be supposed that when they are well developed and actively used by the bird, it metabolic activity in the organism enhanced. An active metabolism implies a great demand for energy. It is thus not surprising to find, in these healthy birds, essential metals present in the soluble fraction where they are more available.

Hepatic Cu was mainly present in S1 where its levels were very significantly correlated to the MT level in the White-Tailed Tropicbird. In this bird, the levels and proportion of hepatic Cu in S1 were the highest of the three studied species. The correlation between Cu and MTs observed in liver may be the sign of the regulation of this element in the tropicbird. In contrast, regulation in the other two species may be less efficient. Further studies are needed to verify this last hypothesis.

Renal Zn was found essentially in the insoluble fraction of the cell in the three seabirds, whereas hepatic Zn’s distribution differed among species. Audubon’s Shearwater accumulated hepatic Zn preferentially in the insoluble fraction, whereas in Barau’s Petrel and the White-Tailed Tropicbird the accumulation was slightly oriented toward the soluble fraction. Audubon’s Shearwater was characterized by the lowest levels of MTs and of most elements. Further investigations are needed to understand whether this difference in the accumulation pattern in Audubon’s Shearwater is the result of differences in its storage capacities (e.g., the presence of a smaller amount of ligands in the soluble fraction) or whether these differences result from its diet, which is much richer in fish than the other two species. As a general rule, the subcellular distribution of Zn in the three Reunion species (71% to 81% of the total Zn in the subcellular fraction; Table 1) seems to be less oriented toward the soluble fraction than in other aquatic birds (Nam et al. 2005). Furthermore, renal Mn and Zn levels were higher in the insoluble fraction of incubating birds in relation to birds presenting no incubation patch. These differences in Mn and Zn accumulation may be due to the particular physiologic state of incubating birds, such as temporarily modified nutritional patterns.

Cd, Hg, and Zn were found to be positively correlated in the seabirds from Reunion Island (data not shown) and had a tendency of increasing in the soluble fraction with the increase of their whole-tissue levels (Figs. 2 and 3). Although Zn is considered the main inducer of MT synthesis, MTs have greater affinities for Cd, Cu, and Hg than for Zn. As such, Cd, Cu, and Hg ions are expected to displace Zn ions from Zn-binding sites through a metal-metal exchange reaction (Eaton et al. 1980). The displaced Zn is then available for initiating MT induction (Roessjadj 1996).

The induction of MT synthesis is known to be associated with an increase of the metal resistance of animals (Eaton et al. 1980; Duncan and Klaverkamp 1983). Indeed, MTs main known functions are (1) the homeostatic regulation of intracellular metals by the binding of excess metals that enter the cell and (2) a “rescue” function for structures impaired by inappropriate metal-binding (Hamilton and Mehrle 1986). The correlations observed between MTs and metal levels in these species suggest that MTs could act as a reservoir for toxic Cd and Hg or for excess Cu and Zn. Correlation between MT levels and molCdCuHgZn, and between MT and Zn levels, were systematically stronger (higher r values) than correlations between levels of MTs and Cd, Cu, or Hg considered separately (Table 3). Consequently, we may conclude that MTs seem to be most often bound (1) exclusively to one or several Zn ion (s) or (2) simultaneously to Cd, Cu Hg, and Zn. Furthermore, not only was Cd positively correlated with MT levels in the three species, it was mostly present in the soluble fraction of the cell, where the large majority of the MT burden is found (Table 1). Cd-induced renal toxicity is thought to be associated with Cd not bound to MTs (Chan 1998). In this study, the sampled birds died from poaching, collisions, or other types of accidents, not indicating fatal intoxication. It can thus be suspected that most of the Cd in S1 was bound to MTs, supporting the hypothesis of a MT-mediated detoxification of Cd. However, MT–Cd compounds seemed unequally distributed between liver and kidney. As noted earlier, MT levels were higher in adult than juvenile birds in kidney but not in liver. Interestingly, this does not correspond to Cd trends in which this nonessential metal was more concentrated in adult than in juvenile birds in kidney as well as liver (Kojadinovic et al. 2007). Cd found in liver may be proportionally less bound to MTs compared with renal Cd. Cd can exit liver in the form of Cd–MT through the blood stream and accumulate in kidney (Chan et al. 1993), suggesting that the liver acts as a “transition organ” for Cd–MT and the kidney as a “storage organ” for Cd–MT.
Hg levels in the storage organs of adult Barau’s Petrels (the most Hg-enriched of the three species) were very high. However, histologic examinations of liver and kidney conducted in seabirds with similar Hg levels to those found in Barau’s Petrel (21 ± 28 vs. 24 ± 14 μg.g⁻¹dw) failed to reveal indications of tissue damage associated with these Hg levels (Elliott et al. 1992). This might be related to efficient Hg detoxification processes in the studied seabirds. Other than the detoxification of Cd, MTs could also be suspected to play a part in the detoxification of Hg. This is, however, difficult to prove because Hg subcellular distribution was not determined, and MT–Hg correlation in Barau’s Petrel was weak. Moreover, the interaction between Hg and Se is considered as the main protective process against Hg toxicity (Parizek and Ostadalova 1967; Magos et al. 1987; Elliott et al. 1992, Kim et al. 1996). Indeed, Hg is demethylated in the presence of Se and stored as nontoxic nonbiodegradable, insoluble tiemannite granules in the liver of certain marine mammals and birds (Koeman et al. 1973; Nigro et al. 2002; Ikemoto et al. 2004). A 1:1 molar ratio between Hg and Se in the liver of seabirds strongly suggests the existence of mercuric selenide (HgSe) (Martorza and Berry 1980; Kim et al 1996; Decataldo et al. 2004).

Such ratios were not found in the Reunion seabirds. Relatively low exposure to Hg may be the reason for the apparent absence of HgSe granules in the insoluble fraction, which was, in addition, confirmed by little Se accumulation in the insoluble fraction. A threshold level of 100 μg g⁻¹dw Hg was noted in the Black-Footed Albatross (Diomedea nigripes), a level above which the detoxification of Hg by Se was triggered (Kim et al. 1996). Hg burdens in Reunion adult seabirds were well under this threshold level with hepatic concentrations of 24.3 μg.g⁻¹dw in petrels, 1.72 μg.g⁻¹dw in shearwaters, and 1.89 μg.g⁻¹dw in the White-Tailed Tropicbirds (Kojadinovic et al. 2007). In these birds, Se was chiefly compartmentalized in the soluble fraction of the cells (Table 1), and Se levels in S1 increased with levels in the whole-liver tissue (Fig. 2). Although Hg subcellular distribution was not studied, significant correlations between Hg and Se suggest that it has a similar distribution pattern. Nam et al. (2005) interpreted such results as an indication of the formation of Hg–Se complexes bound to thiol-containing glutathione molecules (Zilmer et al. 2005) in the soluble fraction. These compounds would act as the precursor of the mineralization of HgSe in lysosomes. Further investigations on total Hg and methyl–mercury (MeHg) distribution in the subcellular fractions should be carried out in liver and kidney of these three species to substantiate these hypothesis.

Conclusion

Many free-living seabirds exhibit relatively high concentrations of trace elements, Cd and Hg in particular, with apparently little or no evident ill effects. This is the case of the birds considered during our study, especially Barau’s Petrel. In the three seabirds, a MT-mediated Cd detoxification process seems to take place. The involvement of MTs in Cu and Zn homeostasis was also confirmed. MT synthesis can be induced by a variety of factors, such as various conditions of physical and physiologic stress, including starvation (Nordberg 1998; Debacker et al. 2001). The study of MC and body weight of these seabirds confirmed this statement. They should thus be taken into account when comparing MT data. Furthermore, it appears as though a Se–Hg detoxifying interaction exists in the two Procellariiform species.

As a whole, this study showed that trace element availability and detoxification were mainly influenced by the same factors that had an important impact on trace element levels in these birds (Kojadinovic et al. 2007): age, diet, and phylogeny. Trace element and MT levels in the three species decreased in the following order: Barau’s Petrel, White-Tailed Tropicbird, and Audubon’s Shearwater. This order follows a gradient of (1) weight of the bird (mean weights of adult birds were respectively, 306, 276, and 197g), (2) percentage of cephalopods in their diets (respectively, 98%, 80%, and 50%), and distance of their fishing zones from the coast during the breeding period. However, White-Tailed Tropicbird stood out in various ways, such as the strategies developed by the fledgling to cope with fasting (Kojadinovic et al. 2007), higher percentages of every element in the soluble fraction of its cells, its efficient Cu regulation, the apparent absence of Se–Hg detoxifying interaction and the potential excretion of Se through eggs in female birds (Kojadinovic et al. 2007). In the light of these results, the Tropicbird’s particularities may be linked to the phylogenetic distance that separates this Phaethonitidae from the two Procellariidae species.

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References


Mason AZ, Jenkins KD (1995) Metal detoxification in aquatic organisms. In: Metal speciation and bioavailability in aquatic system IUPAC series on analytic and physical chemistry of environmental systems. 3. Chichester, UK, John Wiley and Sons (eds), pp 479–607


